

414 Rec'd PCT/PTO 01 MAR 2001

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

0425-0825P

U.S. APPLICATION NO. (If known, see 37 CFR 1.5)

09/786060
NEW

INTERNATIONAL APPLICATION NO.

PCT/JP99/04615

INTERNATIONAL FILING DATE

August 26, 1999

PRIORITY DATE CLAIMED

September 1, 1998

TITLE OF INVENTION

EVALUATION METHOD AND PREPARATION PROCESS OF INJECTION CONTAINING A LIPID A ANALOG

APPLICANT(S) FOR DO/EO/US

KANEKO, Kazuhiro; WATANABE, Tomohiro; ASAI, Yasuyuki; SANO, Yoshihisa; KIKUCHI, Kiyomi; *

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39 (1).
4. ☒ The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☒ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☒ has been transmitted by the International Bureau. WO 00/13029
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☒ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
 - a. ☒ is transmitted herewith.
 - b. ☐ has been previously submitted under 35 U.S.C. 154(d)(4)
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)).
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made.
8. ☐ An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. ☐ An English language translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11. to 20. below concern document(s) or information included:

11. ☒ An Information Disclosure Statement under 37 CFR 1.97 and 1.98-International Search Report (PCT/ISA/210)
12. ☒ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☒ A **FIRST** preliminary amendment.
14. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
15. ☐ A substitute specification.
16. ☐ A change of power of attorney and/or address letter.
17. ☐ A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821-1.825.
18. ☐ A second copy of the published international application under 35 U.S.C. 154(d)(4).
19. ☐ A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
20. ☒ Other items or information:
 - 1.) International Preliminary Examination Report (PCT/IPEA/409)
 - 2.) Four (4) sheets of Formal Drawings

*KUSHIDA, Ikuo; ASHIZAWA, Kazuhide

U.S. APPLICATION NO (if known, see 37 CFR 1.5)

09786060

INTERNATIONAL APPLICATION NO

PCT/JP99/04615

ATTORNEY'S DOCKET NUMBER

0425-0825P

21. ☒ The following fees are submitted:**BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(5):**

Neither international preliminary examination fee (37 CFR 1.482)

nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO

and International Search Report not prepared by the EPO or JPO. \$1,000.00

International preliminary examination fee (37 CFR 1.482) not paid to

USPTO but International Search Report prepared by the EPO or JPO \$860.00

International preliminary examination fee (37 CFR 1.482) not paid to USPTO

but international search fee (37 CFR 1.445(a)(2)) paid to USPTO. \$710.00

International preliminary examination fee (37 CFR 1.482) paid to USPTO

but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$690.00

International preliminary examination fee (37 CFR 1.482) paid to USPTO

and all claims satisfied provisions of PCT Article 33(1)-(4). \$100.00

ENTER APPROPRIATE BASIC FEE AMOUNT =Surcharge of \$130.00 for furnishing the oath or declaration later than ☐ 20 ☐ 30
months from the earliest claimed priority date (37 CFR 1.492(e)).

\$ 0

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE
Total Claims	12 - 20 =	0	X \$18.00
Independent Claims	1 - 3 =	0	X \$80.00
MULTIPLE DEPENDENT CLAIM(S) (if applicable)		None	+ \$270.00

\$ 0

\$ 0

\$ 0

\$ 0

TOTAL OF ABOVE CALCULATIONS = \$ 860.00☐ Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are

reduced by 1/2. \$ 0

SUBTOTAL = \$ 860.00Processing fee of \$130.00 for furnishing the English translation later than ☐ 20 ☐ 30

months from the earliest claimed priority date (37 CFR 1.492(f)). \$ 0

TOTAL NATIONAL FEE = \$ 860.00

Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be

accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +

TOTAL FEES ENCLOSED = \$ 900.00

Amount to be:

refunded \$

charged \$

a. ☒ A check in the amount of \$ 900.00 to cover the above fees is enclosed.b. ☐ Please charge my Deposit Account. No. _____ in the amount of \$ _____ to cover the above fees.

A duplicate copy of this sheet is enclosed.

c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any

overpayment to Deposit Account No. 02-2448.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

Send all correspondence to:

Birch, Stewart, Kolasch & Birch, LLP or Customer No. 2292

P.O. Box 747

Falls Church, VA 22040-0747

(703)205-8000

Date: March 1, 2001

By

Raymond C. Stewart, #21,066

/cgc

09/786060

JC02 Rec'd PCT/PTO 01 MAR 2001

PATENT
0425-0825P

IN THE U.S. PATENT AND TRADEMARK OFFICE

Applicant: KANEKO, Kazuhiro et al. Conf.:
Int'l. Appl. No.: PCT/JP99/04615
Appl. No.: New Group:
Filed: March 1, 2001 Examiner:
For: EVALUATION METHOD AND PREPARATION
PROCESS OF INJECTION CONTAINING A
LIPID A ANALOG

PRELIMINARY AMENDMENT

BOX PATENT APPLICATION

Assistant Commissioner for Patents
Washington, DC 20231

March 1, 2001

Sir:

The following Preliminary Amendments and Remarks are respectfully submitted in connection with the above-identified application.

AMENDMENTS

IN THE SPECIFICATION:

Please amend the specification as follows:

Before line 1, insert --This application is the national phase under 35 U.S.C. § 371 of PCT International Application No. PCT/JP99/04615 which has an International filing date of August 26, 1999, which designated the United States of America.--

FOR PCT/JP99/04615

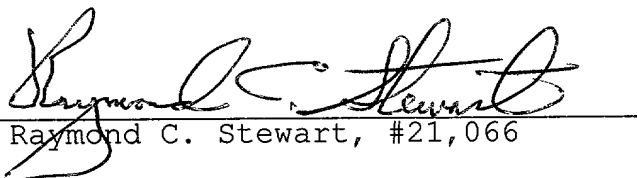
REMARKS

The specification has been amended to provide a cross-reference to the previously filed International Application.

If necessary, the Commissioner is hereby authorized in this, concurrent, and future replies, to charge payment or credit any overpayment to Deposit Account No. 02-2448 for any additional fees required under 37 C.F.R. § 1.16 or under 37 C.F.R. § 1.17; particularly, extension of time fees.

Respectfully submitted,

BIRCH, STEWART, KOLASCH & BIRCH, LLP

By 
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RCS/cqc
0425-0825P

(Rev. 02/12/01)

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09/786060

JC02 Rec'd PCT/PTO 01 MAR 2001

Description

Evaluation method and preparation process of injection containing a lipid A analog

Field of the Invention

The present invention relates to a method for forecasting and evaluating the pharmacokinetics of an injection containing lipid A analog, a quality evaluating method of the injection for ensuring the injection to have constant pharmacokinetics, and a preparation process of the injection.

Prior Art

Lipid A is the main moiety causing activities of lipopolysaccharide (hereinafter referred to as LPS), has various biological activities such as macrophage stimulation, antitumor effect and pyrogenicity (for example, Haruhiko Takada and Shozo Kotani, *Protein, Nucleic Acid and Enzyme*, 31(4), 361 (1986)).

Various lipid A analogs have recently been synthesized and examined for their biological activities (Yuji Ogawa et al., *Metabolism* 26(5), 415 (1989)). Most of the Lipid A analogs having the glycolipid structure are sparingly soluble in water, so that it is difficult to prepare an injection with lipid A analogs.

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In order to prepare an injection and to obtain a highly transparent aqueous solution, the addition of triethylamine, bovine serum albumin, lipids, or the like (Y.B. Kim, et al, *Eur. J. Biochem.* 31, 230 (1972) and R. B. Ramsey, et al, *Blood*, 56, 307 (1980), J. Dijkstra, et al, *J. Immunol.*, 138, 2663 (1987)) and use of basic amino acids or polyamines (JP-A 4-198192) as a solubilizing agent have been reported.

On the other hand, as a method of dispersing a lipid such as lecithin or the like in water to form aggregates of liposomes or the like, known is a method to add a lipid to a buffer having a pH around neutrality, followed by heating and sonication.

The inventors have prepared an injection preparation having a high transparency, which contains aggregates having a diameter not greater than 30 nm, prepared by dissolving the lipid A analog or a pharmacologically acceptable salt thereof produced according to the methods disclosed in JP-A 5-194470 and WO96/39411 in an alkaline aqueous solution and then adding a buffer thereto.

Administration of this injection preparation to the living body of a rat or a beagle is however accompanied with the problem that the blood level of a lipid A analog shows a large variation, depending on the difference in the raw material medicament or preparation lot. This occurs because the state of the aggregates of the medicament

(lipid) in the solution of the injection preparation is not uniform.

It is the common practice to evaluate the state of the aggregates of a lipid in a solution from the appearance observed through an electron microscope, particle size distribution measured by a laser diffraction particle size distribution measuring apparatus or consideration based on the physical properties such as critical micelle concentration or surface tension.

Not so many reports have however been proposed on circular dichroism spectroscopy and/or membrane fluidity evaluation method employed as an evaluation method of the invention.

In circular dichroism spectroscopy (which will hereinafter be abbreviated as "CD spectroscopy"), measurement is carried out reflecting the difference in the refractive index and absorbance of an optically active substance between those relative to left hand circular polarized light and those relative to right hand circular polarized light. This method is frequently employed for the conformation analysis of peptides or proteins or optical activity analysis of low-molecular compounds, but it has hardly been employed for the analysis of lipid-related substances. Reported are only the case wherein a dynamic change of a lipid in a liposome membrane depending on temperature was investigated by the analysis of

variations in a CD spectrum (JP-A 62-252795); the case wherein CD spectroscopy is useful for evaluation of the characteristics of lipid particles in water, is an evaluation method markedly simple in operation and permits measurement of a dynamic state change in the membrane at a diluted concentration (N. Nakashima et al, CHEM. LET., 1503, 10 (1985)); and a case wherein CD spectroscopy is employed as one of the evaluation methods of the correlation between the characteristics of aggregates in various formulations of Prostaglandin E1 in liposome and elution characteristics (Sharon M. K. et al, Biochim. Biophys. Acta, 1327, 97 (1997)).

As a method for evaluating membrane fluidity (softness of membrane), known are fluorescence probe method, electron spin resonance (ESR) method and nuclear magnetic resonance (NMR) method (Robert B Gennith; Biomembrane, p146 (1992), published by Springer Verlag, Tokyo). Among them, the fluorescence probe method is a method for evaluating membrane fluidity having a bimolecular membrane structure of phospholipid. In this method, the state of the membrane in the vicinity of a fluorescent substance is observed by mixing a fluorescence probe such as diphenylhexatriene (which will hereinafter be abbreviated as "DPH") in the membrane of lipid and then, measuring the polarity of fluorescence emitted upon exposure to polarized incident light. There is a report of

the application of this method to a lipid A analog (Braudenburg K, et al., Biochim. Biophys. Acta, 225, 775 (1984)). The fluorescence probe method makes it possible to calculate fluorescence polarity (P: ranging from 0 to 0.5) and/or fluorescence anisotropy (r: ranging from 0 to 0.4) and/or order parameter (S: ranging from 0 to 1.0) by measuring the vertical polarizing component and horizontal polarizing component of fluorescence emitted from a lipid sample (Hiroshi Terada, Tetsuro Yoshimura; Liposome in Life Science (1992), published by Springer Verlag, Tokyo). Herein, as the order parameter (S) approaches 0, the membrane fluidity becomes larger. As it approaches 1, on the other hand, the membrane fluidity becomes smaller.

No report has however been published yet on the definite forecasting or evaluation of the pharmacokinetics of a lipid A analog based on the correlation between the measuring and evaluating results of the aggregate condition of the medicament in a solution and the pharmacokinetics. In addition, a report has been made neither on a process for producing an injection preparation wherein the pharmacokinetics of a lipid A analog has been controlled nor a quality assurance method for ensuring the injection preparation to have predetermined pharmacokinetics, each from the viewpoints of the state of the aggregates of the medicament (lipid) in a solution.

It is possible to prepare an injection preparation

containing aggregates having a diameter not greater than 30 nm by dissolving a lipid A analog or pharmacologically acceptable salt thereof in an aqueous alkaline solution and then adding a buffer thereto. In the resulting injection preparation, the lipid A analog or pharmacologically acceptable salt thereof constitutes endoplasmic reticulum with lipid biomolecular membrane or micelle structure. In other words, an injection of a lipid A analog which has high transparency in the form of an aqueous solution, a preferred pH range as an injection and good stability can be prepared by the above-described process.

The injection preparation thus prepared is however accompanied with the problem that when it is administered to a rat or beagle, the blood level varies largely from one lot to another of a raw material medicament or the preparation. This owes to that the existing state of a lipid A analog in a solution, that is, the aggregate structure in the form of endoplasmic reticulum with lipid biomolecular membrane or micelle having a diameter not greater than 30 nm differs with the lot of the raw material medicament or injection preparation. There is accordingly a strong demand for the development of a practically usable injection of a lipid A analog, that is, an injection having uniform pharmacokinetics, which is typified by the blood level, without being influenced by the difference among the lots of a raw material medicament or injection preparation;

and a forecasting and evaluating method of the pharmacokinetics of the injection.

Disclosure of the Invention

With the foregoing in view, the present inventors have carried out an extensive investigation to search for an injection containing a lipid A analog, having high transparency and having good stability and in addition, exhibiting constant uniform pharmacokinetics free from the influence of variations of a raw material medicament or injection preparation among lots; and forecasting and evaluating method of the pharmacokinetics of the injection. As a result, it has been found that the object can be attained by the below-described constitutions, leading to the completion of the present invention.

In one aspect of the present invention, there is provided a forecasting method of the pharmacokinetics of a lipid A analog in an injection preparation containing the lipid A analog or pharmacologically acceptable salt thereof, which comprises measuring membrane fluidity and/or circular dichroism in a solution.

In another aspect of the invention, there is also provided an evaluation method of the pharmacokinetics of a lipid A analog in an injection preparation containing the lipid A analog or a pharmacologically acceptable salt thereof, which comprises measuring membrane fluidity and/or

circular dichroism in a solution.

In a further aspect of the present invention, there is also provided a quality assurance method of an injection preparation containing a lipid A analog or a pharmacologically acceptable salt thereof, which comprises measuring and evaluating membrane fluidity and/or circular dichroism in a solution, and ensuring the lipid A analog to exhibit constant pharmacokinetics *in vivo*.

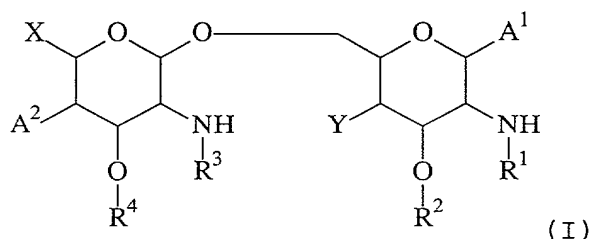
In a still further aspect of the present invention, there is also provided a preparation process of an injection preparation containing a lipid A analog or a pharmacologically acceptable salt thereof, which essentially comprises measuring membrane fluidity and/or circular dichroism in a solution.

The forecasting method of the pharmacokinetics according to the present invention can be employed for the evaluation of an injection preparation, quality evaluation for obtaining an injection preparation exhibiting constant pharmacokinetics or preparation of the injection preparation.

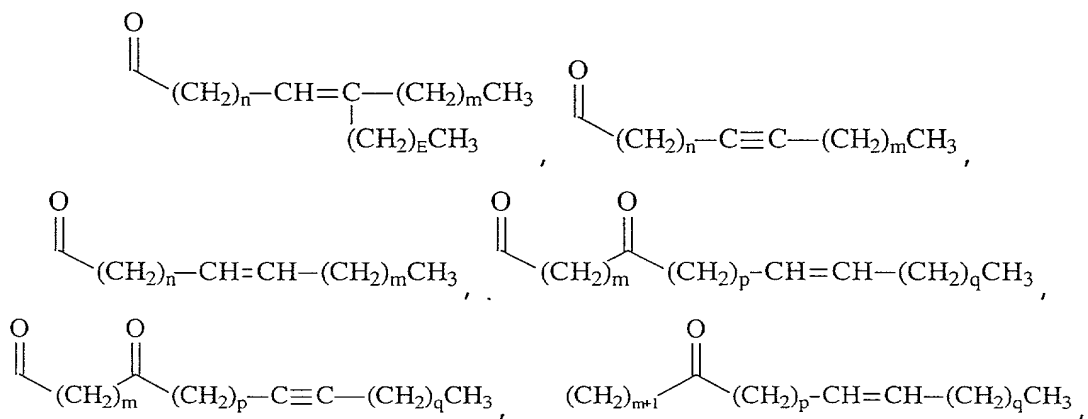
According to the present invention, an injection preparation which is transparent and is ensured to have stable pharmacokinetics is available using a lipid A analog or a pharmacologically acceptable salt (which will hereinafter be called "lipid A analog", collectively). This is an object of the present invention. In addition,

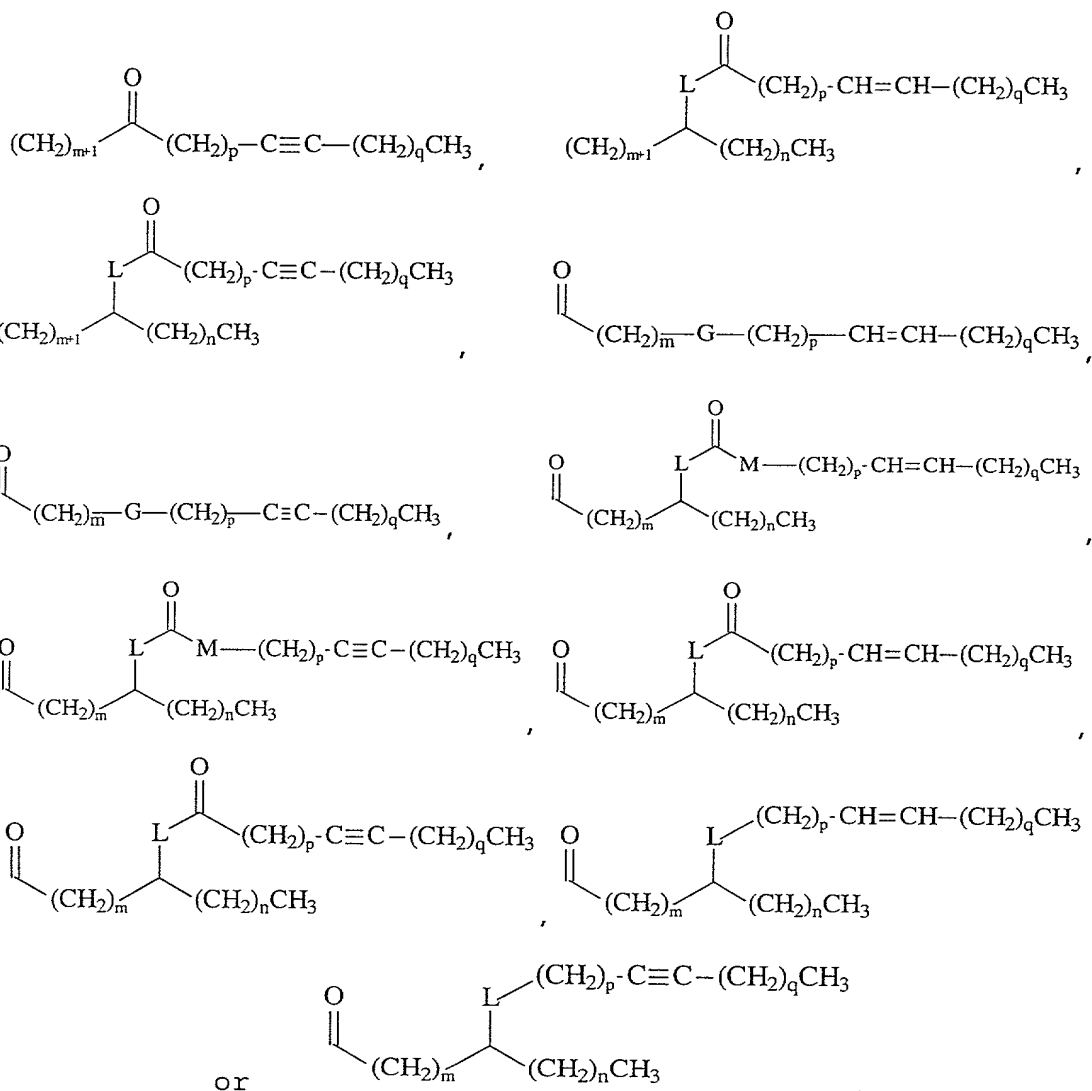
the present invention provides a forecasting and evaluating method of the pharmacokinetics of an injection preparation containing the lipid A analog by measuring the membrane fluidity and/or circular dichroism; and also provides a quality evaluating method of an injection preparation containing a lipid A analog, which ensures the lipid A analog to exhibit constant pharmacokinetics. This is another object of the present invention.

The lipid A analog of the present invention is represented by the following chemical structure, and can be produced by the process disclosed in, for example, JP-A 5-194470 or WO96/39411.

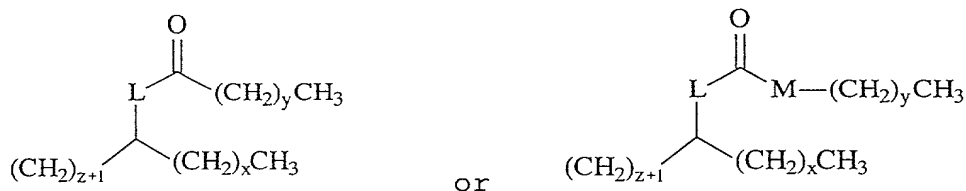
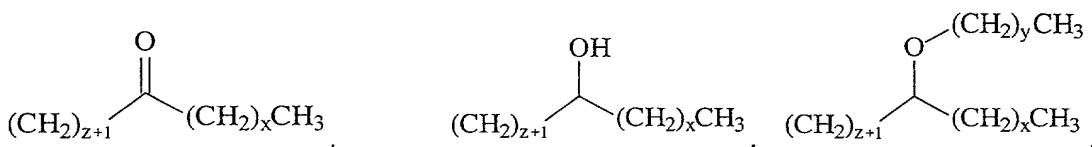
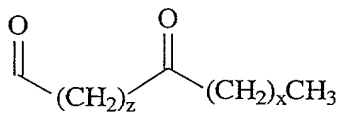
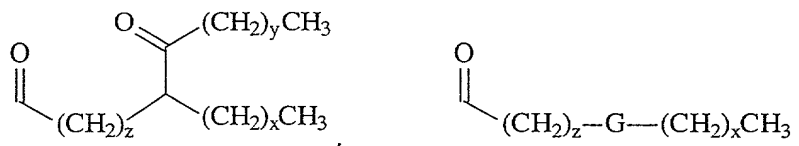
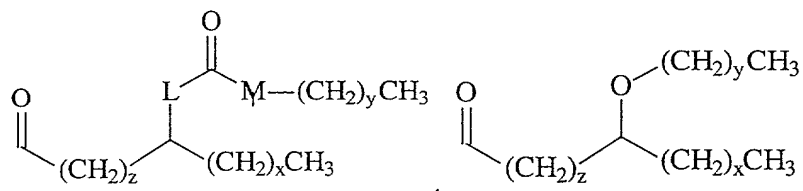
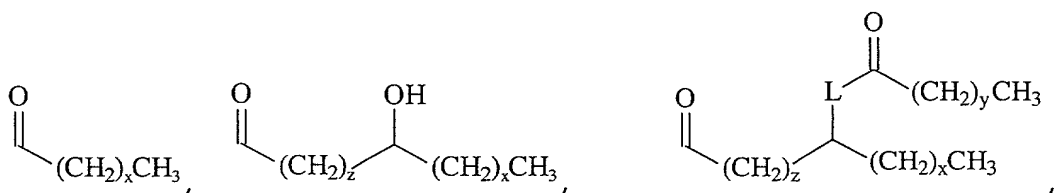


wherein at least one of R^1 , R^2 , R^3 and R^4 is



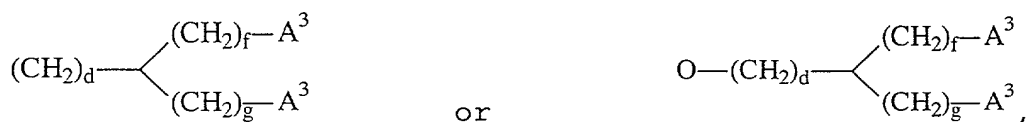
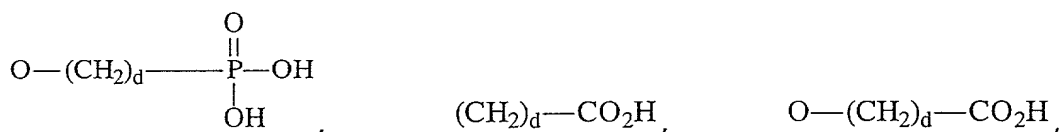
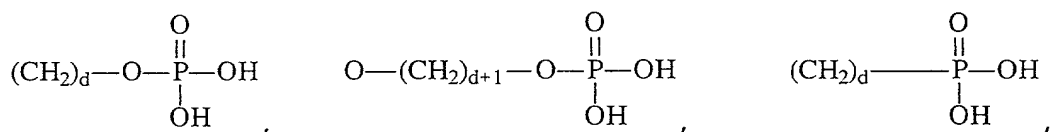


wherein each L is O, N or C; each M is O or N; each E independently is an integer of 0 to 14; each G independently is N, O, S, SO or SO₂; each m independently is an integer of 0 to 14; each n independently is an integer of 0 to 14; each p independently is an integer of 0 to 10; each q independently is an integer of 0 to 10, the rest of R¹, R², R³ and R⁴ are, independently of one another,

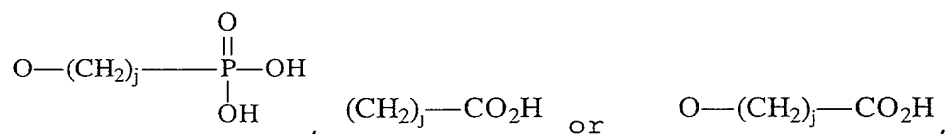
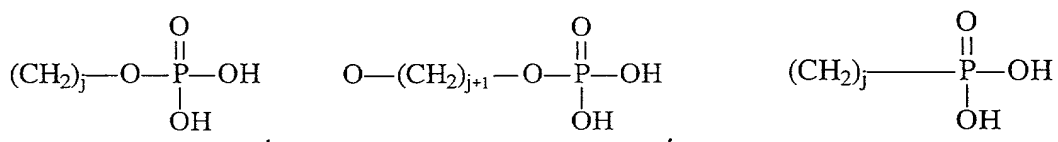


wherein each L is O, N or C; each M is O or N; each x independently is an integer of 0 to 14; each y independently is an integer of 0 to 14; each z independently is an integer of 0 to 10; each G independently is N, O, S, SO or SO₂,

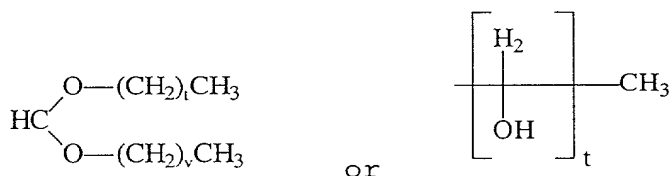
A¹ and A² are, independently of one another, H, OH, OCH₃,



wherein each d independently is an integer of 0 to 5; each f independently is an integer of 0 to 5; each g independently is an integer of 0 to 5; each A³ independently is



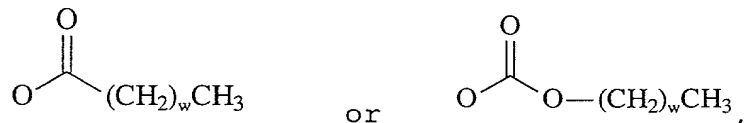
wherein each j independently is an integer of 0 to 14, X is H, (CH₂)_tCH₃, (CH₂)_tOH, (CH₂)_tO(CH₂)_vCH₃, (CH₂)_tOPO(OH)₂, (CH₂)_t-CH=CH-(CH₂)_vCH₃, (CH₂)_t-O-R⁵,



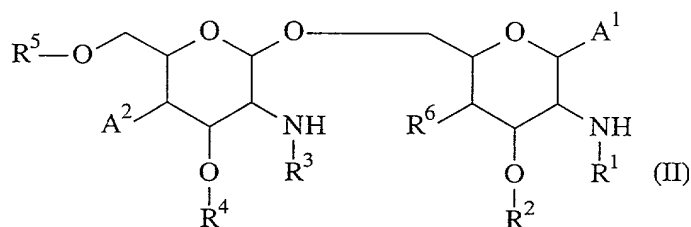
wherein t and v, are independently of one another, an

integer of 0 to 14; R^5 is any of the above definitions of R^1 to R^4 ,

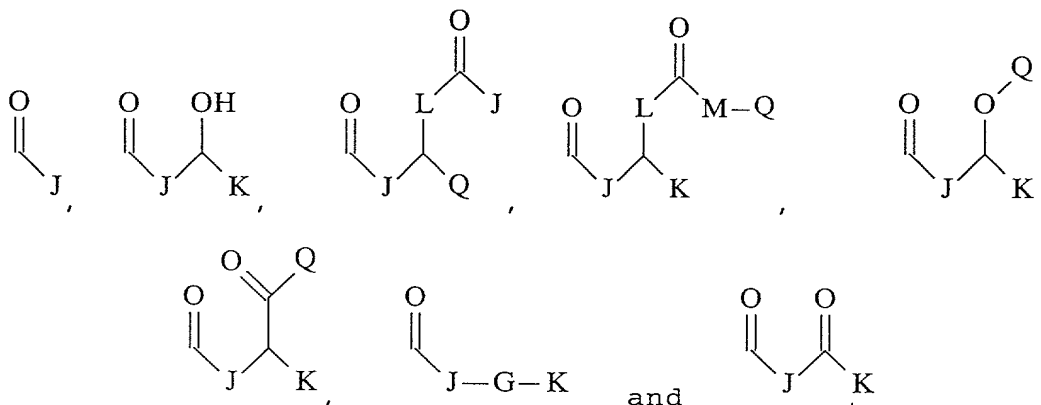
Y is H, OH, $O(CH_2)_wCH_3$, a halogen atom,



wherein w is an integer of 0 to 14.

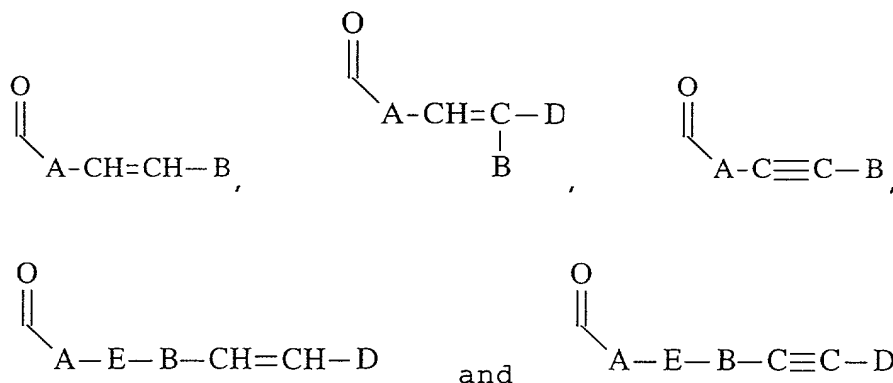


wherein R^1 is a group selected from the groups consisting of

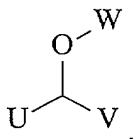


wherein J, K and Q are each a linear or branched alkyl group of 1 to 15 carbon atoms; L is O, NH_2 or CH_2 ; M is O or NH ; G is NH , O, S, SO or SO_2 ,

R^2 is a linear or branched alkyl group of 5 to 15 carbon atoms, R^3 is a group selected from the groups consisting of



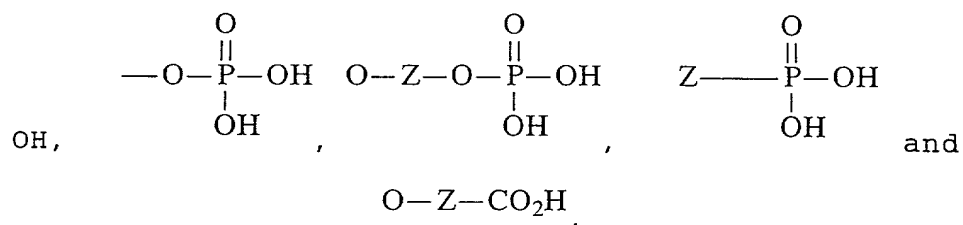
wherein E is N, O, S, SO or SO₂; A, B and D are each a linear or branched alkyl group of 1 to 15 carbon atoms, R⁴ is a group selected from the groups consisting of a linear or branched alkyl group of 4 to 20 carbon atoms and



wherein U and V are each a linear or branched alkyl group of 2 to 15 carbon atoms; W is a hydrogen atom or a linear or branched alkyl group of 1 to 5 carbon atoms,

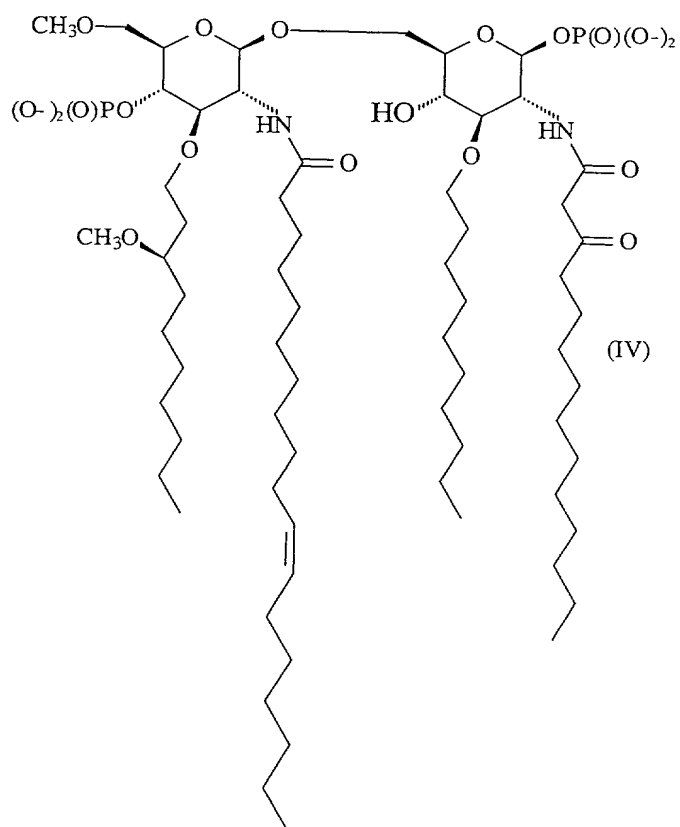
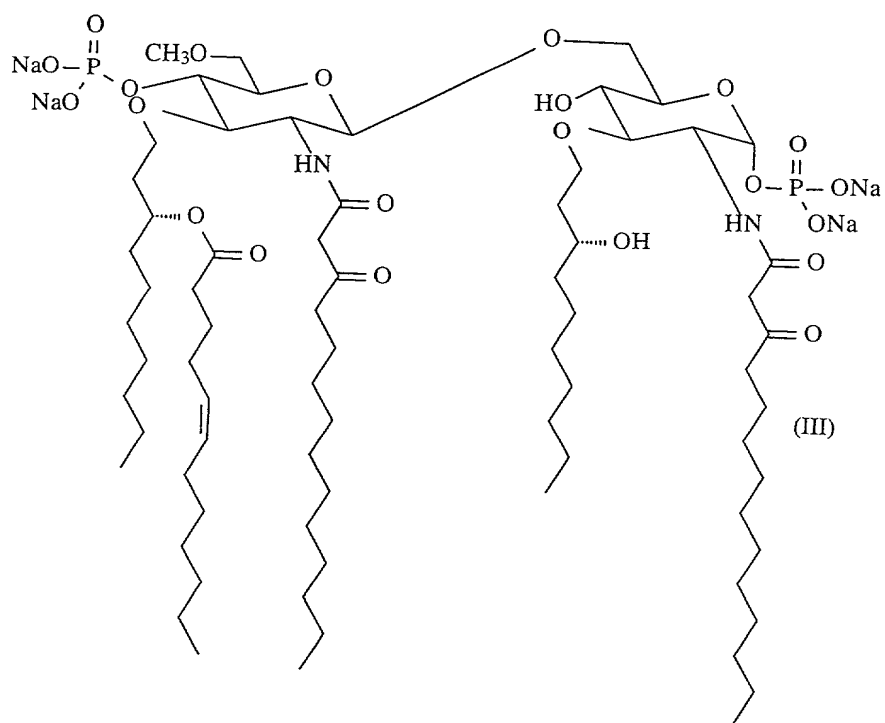
R⁵ is a group selected from the groups consisting of a hydrogen atom, J', -J'-OH, -J'-O-K', -J'-O-K'-OH and -J'-O-PO(OH)₂, wherein J' and K' are each a linear or branched alkyl group of 1 to 5 carbon atoms,

R⁶ is a group selected from the groups consisting of a hydroxyl group, a halogen atom, an alkoxy group of 1 to 5 carbon atoms, and an acyloxy group of 1 to 5 carbon atoms, A¹ and A² are, independently of one another, a group selected from the groups consisting of



wherein Z is a linear or branched alkyl group of 1 to 10 carbon atoms.

Preferred examples of the lipid A analog for use in the present invention include 6-O-[2-deoxy-6-O-methyl-4-O-phosphono-3-O-[(R)-3-Z-dodec-5-enoyloxydecyl]-2-[3-oxotetradecanoylamido]-b-O-phosphono-a-D-glucopyranose] tetrasodium, a-D-glucopyranose, 3-O-decyl-2-deoxy-6-O-[2-deoxy-3-O-(3-methoxydecyl)-6-O-methyl-2-[(1-oxo-11-octadecenyl)amino]-4-O-phosphono-b-D-glucopyranosyl]-2-[(1,3-dioxotetradecyl)amino]-, 1-(dihydrogenphosphate), disodium[6(2Z, 3R)]; and a-D-glucopyranose, 3-O-decyl-2-deoxy-6-O-[2-deoxy-3-O-(3-methoxydecyl)-6-O-methyl-2-[(1-oxo-11-octadecenyl)amino]-4-O-phosphono-b-D-glucopyranosyl]-2-[(1,3-dioxotetradecyl)amino]-, 1-(dihydrogenphosphate), tetrasodium[6(2Z, 3R)]. These compounds are represented by the following chemical structural formulae (III) and (IV).



It is known that the blood level of liposome of phospholipid is influenced largely when it is introduced into a net system such as liver via a scavenger receptor. As a result of study on the mechanism of a scavenger receptor which recognizes and scavenges an anionic high molecule such as liposome, it is also known that membrane fluidity (softness of membrane), surface charge and particle diameter play important roles in the recognition of the scavenger receptor (Hiroshi Terada, Tetsuro Yoshimura; Liposome in Life Science, p326 (1992), published by Springer Verlag, Tokyo).

No example of evaluating physicochemical properties of the membrane of a preparation containing a lipid A analog has so far been proposed from the viewpoint of pharmacokinetics control.

The present invention has revealed that in a step for preparing an injection preparation which contains aggregates having a diameter not greater than 30 nm by dissolving a lipid A analog in an alkaline solution and then adding a buffer to the resulting solution, a pharmacokinetics profile of the injection containing a lipid-A analog can be controlled by measuring and evaluating its membrane fluidity and circular dichroism.

The membrane fluidity of the particles in an injection preparation using DPH has a close correlation with a blood level profile. Upon administration of the

injection preparation, a preparation having larger membrane fluidity (a preparation having softer membrane) disappears more slowly from the blood (larger AUC), while a preparation having smaller membrane fluidity (a preparation having harder membrane) disappears more rapidly from the blood (smaller AUC). This is because after administration of a lipid-A-containing injection, membrane fluidity affects the pharmacokinetics of the medicament. The larger the membrane fluidity, the lipid A does not tend to be trapped easily by a scavenger receptor and is therefore not taken in a phagocyte such as liver cells readily, retarding the disappearance of lipid A from the circulating blood. The present invention has also revealed that there is a correlation between the CD spectrum change and variations in the pharmacokinetics.

The present invention makes it possible to prepare an injection preparation containing a lipid-A analog ensured to have good pharmacokinetics by utilizing, for the measurement of membrane fluidity and/or circular dichroism, the fact that membrane fluidity and/or circular dichroism in the solution of the injection differs definitely between a formulation of a low disappearing rate and a formulation of a high disappearing rate from the blood. The present invention also makes it possible to provide a forecasting or evaluating method of the pharmacokinetics of an injection preparation containing a lipid-A analog by

measuring membrane fluidity and/or circular dichroism; and a quality evaluating method for ensuring the injection to have constant pharmacokinetics.

In the present invention, a highly-transparent injection preparation which has controlled pharmacokinetics and contains aggregates having a diameter not greater than 30 nm can be prepared by dissolving a lipid A analog in an aqueous alkaline solution, adding thereto a buffer and measuring membrane fluidity and/or circular dichroism in the resulting solution. In the present invention, the membrane fluidity of an injection preparation is measured by the fluorescence probe method. This is a method for evaluating membrane fluidity of a bimolecular membrane structure of a phospholipid. Described specifically, the state of the membrane in the vicinity of a fluorescent substance is observed by mixing a fluorescence probe in the membrane of a lipid and measuring the polarity of fluorescence emitted upon exposure to polarized incident light. In the present invention, any one or more than one parameter selected from fluorescence polarity (P: ranging from 0 to 0.5), fluorescence anisotropy (r: ranging from 0 to 0.4) and order parameter (S: ranging from 0 to 1.0) may be used for evaluation of membrane fluidity. As a fluorescence probe to be used, any one capable of emitting stable fluorescence can be employed. Examples include diphenylhexatriene (DPH), carboxyfluorescein, calcein, Nile

Red, pyrene and perylene.

For example, as the order parameter (S) approaches 0, the membrane fluidity is larger, while it approaches 1.0, the membrane fluidity is smaller. The order parameter (S) of the lipid A analog represented by the formula (III) or (IV) is usually 0.5 to 0.7, preferably 0.1 to 0.6, more preferably 0.1 to 0.5.

The circular dichroism spectroscopy (CD spectroscopy) of an injection preparation employed in the present invention is useful as a method for controlling or forecasting the pharmacokinetics of a lipid A analog. It is preferred to evaluate the CD spectrum of each of a number of injection preparations and select the wavelength at which a large difference in CD intensity can be recognized. In the CD spectroscopy of the lipid A analog represented by the formula (III) or (IV), wavelength measured is usually 260 to 320 nm, preferably 270 to 310 nm, more preferably 280 to 300 nm.

As the alkaline aqueous solution in the present invention, the hydroxide of an alkali metal such as sodium hydroxide and potassium hydroxide, preferably sodium hydroxide may be used. The concentration thereof generally ranges from 0.0001M to 0.1M, preferably from 0.0005M to 0.01M and more preferably from 0.001 M to 0.01 M.

In the present invention, after an alkaline aqueous solution is added to a lipid A analog, the temperature of

the mixture may be elevated. The elevated temperature must be higher than the phase transition temperature of the lipid A analog or a pharmacologically acceptable salt thereof, but has no more limitation. The elevated temperature generally ranges from 30°C to 60°C, preferably 45°C to 55°C. The stirring time generally ranges from 10 minutes to 3 hours. Stirring may be carried out with a conventional apparatus. When a lipid A analog is dissolved in a heated alkaline aqueous solution, a lipid A analog may be added to an alkaline aqueous solution which is heated in advance. Alternatively, after a lipid A analog is added to an alkaline aqueous solution, the mixture may be heated. In the present invention, the purpose of elevating the temperature is to accelerate hydration of lipid A analogs to improve dispersibility by elevating the temperature to the phase transition temperature of lipid A analogs or higher, to thereby obtain a transparent solution by stirring for a shortened period of time.

The lipid A analog represented by the formula (IV) however may be heated or stirred at room temperature for 10 minutes to 1.5 hours.

Examples of the component of the buffer for use in the present invention include phosphates, Tris(hydroxymethyl)aminomethane, citrates, glycine and the like. The concentration of the buffer generally ranges from 1 mM to 20 mM. The final pH value of the aqueous

solution of the lipid A analog is preferably 4 to 9, more preferably 6 to 8, still more preferably 6.8 to 7.8. The final pH value may be adjusted by adding a solution of sodium hydroxide, hydrochloric acid, or the like after the addition of the buffer.

If necessary, addition of saccharides and/or amino acids to the buffer may give a more preferable result. In this case, saccharides and/or amino acids to be added may be either one, or two or more kinds thereof. Examples of the saccharide include milk sugar (lactose), sorbitol, glucose, trehalose, mannitol, dextran, and the like. Examples of the amino acid include neutral amino acids such as glycine, acidic amino acids such as aspartic acid, and basic amino acids such as arginine.

In the present invention, the resulting lipid A analog aqueous solution may be freeze-dried by a conventional method to obtain a freeze-dried preparation. Namely, lipid A or an analog thereof is dissolved in an alkaline aqueous solution, further stirred at an elevated temperature if necessary, followed by addition of a buffer to adjust pH of the mixture. After sterilizing filtration, the mixture is filled into a vial or the like, followed by freeze-drying to give a freeze-dried preparation.

When a preparation for injection according to the present invention is administered in the form of aqueous solution, the osmotic pressure ratio of the preparation is

preferably adjusted to a value suitable for administering to humans, generally around 1.

One example of each of the measuring methods of membrane fluidity and circular dichroism (CD spectroscopy) of an injection preparation and the evaluation method of the pharmacokinetics in a rat administered with the preparation will next be described.

Measuring method of membrane fluidity of an injection preparation

It will be evaluated using as a parameter fluorescence polarity (P) and/or fluorescence anisotropy (r) and/or order parameter (S) in accordance with the fluorescence probe method.

First, a solution having a concentration of 0.5 to 0.6 mg/ml is prepared as an injection preparation containing a lipid-A analog (a freeze-dried preparation in a 1.5 to 1.6 mg/vial is prepared by adding 3 ml of distilled water). Then, 4 μ l of a fluorescence probe solution obtained by dissolving 1.5 mg of DPH in 10 ml of THF is added, followed by sufficient mixing. The resulting mixture is allowed to stand at 50°C for 1 hour. After cooling to room temperature, P and/or r of the reaction mixture is measured by a fluorescent spectrophotometer. The measuring conditions of a fluorescent spectrophotometer "Model F-450" (trade name; product of Hitachi Ltd.) are as follows: excited wavelength of 360 nm, luminous wavelength

of 428 nm, applied voltage of 700V, measuring temperature of 25°C and slit width of 5 nm. The order parameter (S) is calculated based on the equation: $S = (r/0.398)^{1/2}$.

It can be judged that membrane fluidity is larger (the membrane is softer) at S closer to 0 and it is smaller (the membrane is harder) at S closer to 1. Measuring method of circular dichroism (CD spectroscopy) of the preparation

A solution having a concentration of 0.5 to 0.6 mg/ml is prepared as an injection containing a lipid A analog (a freeze-dried preparation in a 1.5 to 1.6 mg/vial is prepared by adding 3 ml of distilled water). The CD intensity of this preparation is evaluated by "Spectropolarimeter J720WI" (trade name; product of JASCO Corporation). Measurement is conducted under the conditions of wavelength of 200 to 500 nm (280 nm when fixed wavelength is employed), measuring temperature at room temperature, cell length of 1 cm, scanning speed of 20 nm/min and integration of 5 times. Evaluation method of pharmacokinetics in a rat administered with an injection preparation

The pharmacokinetics profile of the preparation is judged using as a parameter an area under the curve available from the time-dependent curve of the blood level of medicament (this area will hereinafter be abbreviated as "AUC").

First, as an injection preparation containing a lipid A analog, a solution having a concentration of 0.6 mg/ml is prepared by adding distilled water for injection. The resulting solution is administered (0.6 mg/Kg) to a male SD rat in a dose of 1 ml/Kg from its femoral vein. In addition to the blood collection prior to administration, the blood is collected 2, 5, 15, 30, 60 and 120 minutes after administration, each in an amount of 0.25 ml. Each of the bloods thus collected is centrifuged, whereby the plasma is obtained. From the plasma, the lipid A analog is extracted by liquid-liquid extraction, followed by labeling with fluorescence by using an ADAM reagent (product of Funakoshi Co., Ltd.). After solid layer extraction, the concentration of the lipid A analog contained in the plasma is measured using high performance liquid chromatography in accordance with the fluorescence detection method. The AUC is calculated from a change in the blood level from 0 to 120 minutes.

According to the present invention, it is possible to provide an injection preparation containing a lipid A analog as that having high transparency and stably controlled pharmacokinetics. Also a forecasting method and evaluating method of the pharmacokinetics of an injection preparation and a quality assurance method of an injection preparation containing a lipid A analog, which ensures the preparation to have constant pharmacokinetics can be

provided. The following are the advantage examples of the present invention.

Brief Description of the Drawings

Fig. 1 is a graph showing a change in the blood level of a rat administered (0.6mg/kg) with an injection preparation containing the lipid A analog represented by the formula (IV).

Fig. 2 is a graph showing a CD spectrum (spectrum as evaluated by circular dichroism spectroscopy) of an injection preparation containing the lipid A analog represented by the formula (IV).

Fig. 3 is a graph showing a correlation between CD intensity and AUC (upon administration to a rat) of an injection preparation containing the lipid A analog represented by the formula (IV).

Fig. 4 is a graph showing a correlation between order parameter (S) and AUC (upon administration to a rat) of an injection preparation containing the lipid A analog represented by the formula (IV).

Fig. 5 is a graph showing a correlation between the CD intensity and order parameter (S) of an injection preparation containing the lipid A analog represented by the formula (IV).

Fig. 6 is a graph showing a correlation between the order parameter (S) and AUC (upon administration to rat) of

an injection preparation containing the lipid A analog represented by the formula (III).

Fig. 7 is a graph showing a correlation between the particle size and AUC (upon administration to a rat) of an injection preparation containing the lipid A analog represented by the formula (III).

Fig. 8 is a graph showing a correlation between the concentration of sodium hydroxide and AUC (upon administration to a rat) in a dissolution step of an injection preparation containing the lipid A analog represented by the formula (IV).

Test 1

Evaluation of the membrane fluidity and circular dichroism of the preparations different in the profile of pharmacokinetics (AUC), according to the present invention

1) An injection preparation having a small clearance was prepared in the following manner. In a 50 ml glass beaker, 32.47 mg of the lipid A analog represented by the formula (IV) was weighed. After the addition of 7.5 mL of 0.01M NaOH, the resulting mixture was stirred by a stirrer bar (2cm long) at room temperature for 60 minutes. To the reaction mixture was added 25 mL of a lactose phosphate buffer (an aqueous solution obtained by dissolving in 25 mL of purified water 500 mg of lactose monohydrate, 2.25 mg of $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$ and 1.75 mg of $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$). After stirring for

5 minutes by a stirrer bar, the whole amount of the mixture was transferred to a 50 mL measuring flask. Distilled water for injection was then added to adjust the total amount to 50 mL.

2) An injection preparation having a large clearance was prepared in the following manner. In a 50 ml glass beaker, 32.97 mg of the lipid A analog represented by the formula (IV) was weighed. After the addition of 0.25 mL of 0.01M NaOH and 7.25 mL of distilled water for injection, the resulting mixture was stirred by a stirrer bar (2 cm long) at room temperature for 60 minutes. To the reaction mixture was added 25 mL of a lactose phosphate buffer (an aqueous solution obtained by dissolving in 25 mL of purified water 500 mg of lactose monohydrate, 2.25 mg of $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$ and 1.75 mg of $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$). After stirring for 5 minutes by a stirrer bar, the whole amount of the mixture was transferred to a 50 mL measuring flask and then mixed with 7.45 mL of 0.01M NaOH. By the further addition of 0.01M NaOH, pH of the mixture was adjusted to 7.4. Distilled water for injection was then added to increase the total amount to 50 mL.

These two injection preparations thus prepared, that is, a formulation 1) having a small clearance and a formulation 2) having a large clearance, were evaluated for a correlation between AUC and each of membrane fluidity and circular dichroism.

As illustrated in Fig. 1, a change in the blood level of a rat administered with an injection preparation containing a lipid A analog differed largely between an injection preparation of a small clearance and that of a large clearance. The order parameter (S) as an indicator of membrane fluidity was 0.499 in the case of an injection preparation having a small clearance, while that in the case of an injection preparation having a large clearance was 0.624, indicating that there was a significant difference between them.

In CD intensity as measured by circular dichroism (CD spectroscopy), as shown in Fig. 2, a clear difference was recognized between them at wavelength of 280 nm. The above-described findings show clearly that evaluation of membrane fluidity and/or circular dichroism is useful for evaluating a change in the pharmacokinetics of the lipid-A-analog-containing injection preparation according to the present invention.

Test 2

Measurement and evaluation of membrane fluidity and/or circular dichroism of the injection preparation according to the present invention and a correlation between AUC and membrane fluidity or circular dichroism

Ten lots of an injection preparation containing the lipid A analog represented by the formula (IV) were prepared and a correlation between CD intensity (at

wavelength of 280 nm) and/or order parameter (S) and AUC of a rat intravenously administered with the preparation was evaluated. As a result, as shown in Fig. 3, a good, positive correlation was recognized between AUC of the rat and CD intensity. The preparation having a greater CD intensity exhibited greater AUC. As shown in Fig. 4, a good, negative correlation was, on the other hand, recognized between AUC of the rat and order parameter (S). The smaller the order parameter (S) (as it approached 0), the greater AUC. As shown in Fig. 5, a good, negative correlation was recognized between the CD intensity (at wavelength of 280 nm) and order parameter (S) of the injection preparation. The greater the CD intensity, the smaller the order parameter (S) (it approached 0).

Concerning an injection preparation containing the lipid A analog represented by the formula (III), ten lots were prepared and evaluated. As a result, a negative correlation was recognized between the AUC of a rat and order parameter (S) as shown in Fig. 6 and its correlation coefficient was 0.873. Concerning the correlation between AUC of a rat and the average particle diameter of the injection preparation, AUC showed large variations at an average particle diameter ranging from 10 to 20 nm and therefore, no definite correlation was recognized (Fig. 7) between them. Accordingly, evaluation using an average particle size was insufficient for forecasting and

evaluation of pharmacokinetics.

Thus, a good correlation of a pharmacokinetics profile (AUC) with evaluation of membrane fluidity and/or circular dichroism of the injection preparation according to the present invention has been proved. It is evident that use of membrane fluidity (order parameter (S)) and circular dichroism (CD) is fully effective for forecasting and evaluating the pharmacokinetics of an injection preparation containing a lipid A analog. In addition, it becomes possible to prepare an injection preparation having ensured pharmacokinetics by evaluating the state of aggregates in the solution based on membrane fluidity and/or circular dichroism. In other words, AUC can be presumed by measuring the order parameter (S) and/or circular dichroism (CD) of an injection preparation containing lipid A analog. Accordingly, the present invention can therefore be used industrially and useful.

Examples

The present invention will hereinafter be described more specifically by referring to Examples. However, the present invention is not limited by them.

In Examples 1 to 10 which will be described below, shown are aqueous injections prepared by changing the concentration of sodium hydroxide (NaOH) used for dissolving therein a lipid A analog represented by the

formula (IV). Table 1 shows the CD intensity and the order parameter (S) of each injection preparation and AUC upon administration to a rat. Further, in Fig. 8 is shown a correlation between the concentration of sodium hydroxide (NaOH) used in the above-described dissolving step and AUC. The greater the concentration of sodium hydroxide (NaOH), the greater each of AUC and order parameter (S). At 0.001 to 0.01M, AUC and order parameter (S) reached a plateau. This means that the ensured and/or forecast optimum concentration of sodium hydroxide (NaOH) used in the dissolving step of an injection preparation containing the lipid A analog represented by the formula (IV) was 0.001 to 0.01M.

Example 1

In a 50 mL glass beaker, 32.47 mg of a lipid A analog of the formula (IV) was weighed and 7.5 mL of 0.01M NaOH was added to it. The resulting mixture was stirred by a stir bar of 2 cm long at room temperature for 60 minutes (the concentration of sodium hydroxide (NaOH) employed in the dissolving step: 0.01M). To the reaction mixture was added 25 mL of a lactose phosphate buffer (25 mL of an aqueous solution having 500 mg of lactose monohydrate, 2.25 mg of $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$ and 1.75 mg of $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$ dissolved therein). After stirring by a stirrer for 5 minutes, the total amount of the reaction mixture was transferred to a 50 mL measuring flask. Distilled water for injection was

then added to adjust the total amount to 50 mL. The solution at this time had a pH of 7.39.

Example 2

In a 50 mL glass beaker, 2.23 mg of the lipid A analog of the formula (IV) was weighed and 0.75 mL of 0.01M NaOH and 6.75 mL of distilled water for injection were added to it. The resulting mixture was stirred by a stir bar of 2cm long at room temperature for 60 minutes (the concentration of sodium hydroxide (NaOH) employed in the dissolving step: $0.01M \times 1/10$). To the reaction mixture was added 25 mL of a lactose phosphate buffer (a 25 mL aqueous solution having 500 mg of lactose monohydrate, 2.25 mg of $Na_2HPO_4 \cdot 7H_2O$ and 1.75 mg of $NaH_2PO_4 \cdot H_2O$ dissolved therein). After stirring by a stirrer for 5 minutes, the total amount of the reaction mixture was transferred to a 50 mL measuring flask. In the measuring flask, 6.75 mL of 0.01M NaOH was charged and they were mixed. Distilled water for injection was then added to adjust the total amount to 50 mL. The solution at this time had a pH of 7.41.

Example 3

In a 50mL glass beaker, 32.13mg of the lipid A analog of the formula (IV) was weighed and 0.25mL of 0.01N NaOH and 7.25mL of distilled water for injection were added to it. The resulting mixture was stirred by a stir bar of 2cm long at room temperature for 60 minutes (the concentration

of sodium hydroxide (NaOH) employed in the dissolving step: 0.01M x 1/30). To the reaction mixture was added 25mL of a lactose phosphate buffer (a 25mL aqueous solution having 500mg of lactose monohydrate, 2.25mg of $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$ and 1.75mg of $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$ dissolved therein). After stirring by a stirrer for 5 minutes, the total amount of the reaction mixture was transferred to a 50mL measuring flask. In the measuring flask, 7.25mL of 0.01M NaOH was charged further and they were mixed. Distilled water for injection was then added to adjust the total amount to 50mL. The solution at this time had a pH of 7.42.

Example 4

In a 50 mL glass beaker, 32.16 mg of the lipid A analog of the formula (IV) was weighed and 0.125 mL of 0.01M NaOH and 7.375 mL of distilled water for injection were added to it. The resulting mixture was stirred by a stirbar of 2cm long at room temperature for 60 minutes (the concentration of sodium hydroxide (NaOH) employed in the dissolving step: 0.01M x 1/60). To the reaction mixture was added 25 mL of a lactose phosphate buffer (a 25 mL aqueous solution having 500 mg of lactose monohydrate, 2.25 mg of $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$ and 1.75 mg of $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$ dissolved therein). After stirring by a stirrer for 5 minutes, the total amount of the reaction mixture was transferred to a 50 mL measuring flask. In the measuring flask, 7.375 mL of 0.01M NaOH was charged further and they were mixed.

Distilled water for injection was then added to adjust the total amount to 50 mL. The solution at this time had a pH of 7.30.

Example 5

In a 50 mL glass beaker, 32.63 mg of the lipid A analog of the formula (IV) was weighed. A 5.0 mL portion of the mixture of 0.125 mL of 0.01M NaOH and 7.375 mL of distilled water for injection was added, followed by further addition of 2.5 mL of distilled water for injection. The resulting mixture was stirred by a stir bar of 2cm long at room temperature for 60 minutes (the concentration of sodium hydroxide (NaOH) employed in the dissolving step: $0.01M \times 1/90$). To the reaction mixture was added 25 mL of a lactose phosphate buffer (a 25 mL aqueous solution having 500 mg of lactose monohydrate, 2.25 mg of $Na_2HPO_4 \cdot 7H_2O$ and 1.75 mg of $NaH_2PO_4 \cdot H_2O$ dissolved therein). After stirring by a stirrer for 5 minutes, the total amount of the reaction mixture was transferred to a 50 mL measuring flask. In the measuring flask, 7.4 mL of 0.01M-NaOH was charged. After mixing, 0.01M NaOH was added further to adjust the pH of the mixture to 7.4. Distilled water for injection was then added to give the total amount of 50 mL. The solution at this time had a pH of 7.42.

Example 6

In a 50 mL glass beaker, 32.07 mg of the lipid A analog of the formula (IV) was weighed, followed by the

addition of a 3.75 mL portion of a solution containing 0.125 mL of 0.01M NaOH and 7.375 mL of distilled water for injection. To the resulting mixture, 3.75 mL of distilled water for injection was added further. The resulting mixture was stirred by a stir bar of 2cm long at room temperature for 60 minutes (the concentration of sodium hydroxide (NaOH) employed in the dissolving step: $0.01M \times 1/120$). To the reaction mixture was added 25 mL of a lactose phosphate buffer (a 25 mL aqueous solution having 500 mg of lactose monohydrate, 2.25 mg of $Na_2HPO_4 \cdot 7H_2O$ and 1.75 mg of $NaH_2PO_4 \cdot H_2O$ dissolved therein). After stirring by a stirrer for 5 minutes, the whole amount of the reaction mixture was transferred to a 50 mL measuring flask. In the measuring flask, 7.4 mL of 0.01N NaOH was further charged and they were mixed. The mixture was adjusted to pH 7.4 by further addition of 0.01M NaOH. Distilled water for injection was then added to give the total amount of 50 mL. The solution at this time had a pH of 7.48.

Example 7

In a 50 mL glass beaker, 32.43 mg of the lipid A analog of the formula (IV) was weighed. A 5.0 ml portion of a solution obtained by mixing 0.125 mL of 0.01M NaOH and 14.875 mL of distilled water for injection was then added, followed by the addition of 2.5 mL of distilled water for injection. The resulting mixture was stirred by a stir bar

of 2cm long at room temperature for 60 minutes (the concentration of sodium hydroxide (NaOH) employed in the dissolving step: $0.01M \times 1/180$). To the reaction mixture was added 25 mL of a lactose phosphate buffer (a 25 mL aqueous solution having 500 mg of lactose monohydrate, 2.25 mg of $Na_2HPO_4 \cdot 7H_2O$ and 1.75 mg of $NaH_2PO_4 \cdot H_2O$ dissolved therein). After stirring by a stirrer for 5 minutes, the whole amount of the reaction mixture was transferred to a 50 mL measuring flask. In the measuring flask, 7.45 mL of 0.01M-NaOH was further charged and they were mixed. After pH adjustment to 7.4 with 0.01M NaOH, distilled water for injection was added to give the total amount of 50 mL. The solution at this time had a pH of 7.41.

Example 8

In a 50 mL glass beaker, 32.84 mg of the lipid A analog of the formula (IV) was weighed. A 3.75 ml portion of the solution obtained by mixing 0.125 mL of 0.01M NaOH and 14.875 mL of distilled water for injection was then added, followed by the addition of 3.75 mL of distilled water for injection. The resulting mixture was stirred by a stir bar of 2cm long at room temperature for 60 minutes (the concentration of sodium hydroxide (NaOH) employed in the dissolving step: $0.01M \times 1/240$). To the reaction mixture was added 25 mL of a lactose phosphate buffer (a 25 mL aqueous solution having 500 mg of lactose monohydrate, 2.25 mg of $Na_2HPO_4 \cdot 7H_2O$ and 1.75 mg of $NaH_2PO_4 \cdot H_2O$ dissolved

therein). After stirring by a stirrer for 5 minutes, the whole amount of the reaction mixture was transferred to a 50 mL measuring flask. In the measuring flask, 7.45 mL of 0.01M NaOH was charged and they were mixed. After pH adjustment to 7.4 by the addition of 0.01M-NaOH, distilled water for injection was added to give the total amount of 50 mL. The solution at this time had a pH of 7.41.

Example 9

In a 50 mL glass beaker, 32.35 mg of a lipid A analog of the formula (IV) was weighed. A 0.75 ml portion of the solution obtained by mixing 0.25 mL of 0.01M NaOH and 7.25 mL of distilled water for injection was then added, followed by the addition of 6.75 mL of distilled water for injection. The resulting mixture was stirred by a stir bar of 2cm long at room temperature for 60 minutes (the concentration of sodium hydroxide (NaOH) employed in the dissolving step: $0.01M \times 1/350$). To the reaction mixture was added 25 mL of a lactose phosphate buffer (a 25 mL aqueous solution having 500 mg of lactose monohydrate, 2.25 mg of $Na_2HPO_4 \cdot 7H_2O$ and 1.75 mg of $NaH_2PO_4 \cdot H_2O$ dissolved therein). After stirring by a stirrer for 5 minutes, the whole amount of the reaction mixture was transferred to a 50 mL measuring flask. In the measuring flask, 7.45 mL of 0.01M-NaOH was charged and they were mixed. After pH adjustment with 0.01M NaOH to 7.4, distilled water for injection was added to give the total amount of 50 mL. The

solution at this time had a pH of 7.41.

Example 10

In a 50 mL glass beaker, 32.97 mg of the lipid A analog of the formula (IV) was weighed. A 0.5 ml portion of a solution obtained by mixing 0.25 mL of 0.01M NaOH and 7.25 mL of distilled water for injection was then added, followed by the addition of 7.0 mL of distilled water for injection. The resulting mixture was stirred by a stir bar of 2cm long at room temperature for 60 minutes (the concentration of sodium hydroxide (NaOH) employed in the dissolving step: $0.01M \times 1/450$). To the reaction mixture was added 25 mL of a lactose phosphate buffer (a 25 mL aqueous solution having 500 mg of lactose monohydrate, 2.25 mg of $Na_2HPO_4 \cdot 7H_2O$ and 1.75 mg of $NaH_2PO_4 \cdot H_2O$ dissolved therein). After stirring by a stirrer for 5 minutes, the whole amount of the reaction mixture was transferred to a 50 mL measuring flask. In the measuring flask, 7.45 mL of 0.01M NaOH was charged and they were mixed. After pH adjustment with 0.01M-NaOH to 7.4, distilled water for injection was added to give the total amount of 50 mL. The solution at this time had a pH of 7.43.

Table 1: Physicochemical parameter of an injection preparation containing a lipid A analog and AUC (0-2hr) of a rat administered with the preparation

Example	Concentration (M) of NaOH in the dissolving step	AUC (0-2hr) (ng·hr/ml)	CD Intensity	Order parameter (S)
Ex 1	0.01	7377±261	-14.1	0.499
Ex 2	0.01×1/10	7465±276	-13.9	0.491
Ex 3	0.01×1/30	6231±318	-17.7	0.540
Ex 4	0.01×1/60	5817±117	-18.6	0.553
Ex 5	0.01×1/90	5158±173	-20.5	0.584
Ex 6	0.01×1/120	5078±290	-22.1	0.595
Ex 7	0.01×1/180	4388±358	-22.3	0.608
Ex 8	0.01×1/240	1665±106	-24.5	0.650
Ex 9	0.01×1/300	3858±255	-23.1	0.624
Ex 10	0.01×1/450	-	-21.9	0.616

AUC: Mean±S.D.

In Examples 11 to 21 which will be described below, stirring was conducted at room temperature in Examples 11 to 16, while ultrasonic dispersion (at 10°C or less) was conducted in Examples 17 to 21, each in the dissolving step for the preparation of an injection preparation containing the lipid A analog represented by the formula (III). Since the phase transition temperature of the lipid A analog represented by the formula (III) is about 30°C, sufficient dissolution and dispersion was not available in Examples 11 to 21 and large variations in AUC of the rat administered with the preparation and physicochemical parameters including order parameter (S) were recognized as shown in Table 2.

Examples 11 to 16

In a glass beaker, 100 mg of the lipid A analog represented by the formula (III) was weighed, followed by the addition of 50 mL of 0.01M NaOH. The resulting mixture was stirred by a stirrer at room temperature (about 25°C). To the reaction mixture was added 600 mL of a lactose phosphate buffer (an aqueous solution obtained by dissolving in 600 mL of distilled water for injection 100 g of lactose monohydrate, 0.45 g of $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$ and 0.35 g of $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$). After stirring by a stirrer, a proper amount of distilled water for injection was added to adjust the total amount to 1 L. The medicament solution thus obtained was filtered through a 0.22 μm filter. Then, 5.3 ml portions of the filtrate were pipetted into vials, followed by freeze-drying. To each of the vials containing the freeze-dried injection preparation, 5 ml of distilled water for injection was added to thaw the preparation. The evaluation results of the pharmacokinetics (AUC) of a rat, particle size, and membrane fluidity (order parameter (S), fluorescence anisotropy r, fluorescence polarity P) are shown in Table 2.

Example 17-21

In a glass beaker, 100 mg of the lipid A analog represented by the formula (III) was weighed, followed by the addition of 150 mL of 0.01M NaOH. The resulting mixture was stirred by a stirrer at room temperature. After visual conformation of the disappearance of a gel of

the lipid analog A represented by the formula (III), the glass beaker was subjected to ultrasonic exposure at a temperature maintained at 10°C or less in a bath type sonicator. To the reaction mixture was added 600 mL of a lactose-phosphate buffer (an aqueous solution obtained by dissolving in 600 mL of distilled water for injection 100 g of lactose monohydrate, 0.45 g of $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$ and 0.35 g of $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$). After stirring by a stirrer, a proper amount of distilled water for injection was added to adjust the total amount to 1 L. The medicament solution thus obtained was filtered through a 0.22 μm filter. Then, 5.3 ml portions of the filtrate were pipetted into vials, followed by freeze-drying. To each of the vials containing the freeze-dried injection preparation, 5 mL of distilled water for injection was added to thaw the preparation. The evaluation results of the pharmacokinetics (AUC) of a rat, particle diameter, and membrane fluidity (order parameter (S), fluorescence anisotropy r, fluorescence polarity P) are shown in Table 2.

Table 2

Physicochemical parameters of an injection preparation containing a lipid A analog ($500 \mu\text{g/vial}$) and AUC (0-2hr) of a rat administered with the preparation

Example	Concentration (M) of NaOH in the dissolving step	Dispersing method	AUC (0-2hr) (ng·hr/ml)	Particle diameter (nm)	fluorescence anisotropy (r)	fluorescence polarity (p)	Order parameter (S)
Ex 11	0.01	Stirring under heat	2762	33.0 ± 9.9	0.215	0.291	0.735
Ex 12	0.01	Stirring under heat	1130	40.1 ± 13.0	0.23	0.310	0.760
Ex 13	0.01	Stirring under heat	624	60.9 ± 16.9	0.242	0.324	0.780
Ex 14	0.01	Stirring under heat	2007	30.7 ± 11.3	0.201	0.274	0.711
Ex 15	0.01	Stirring under heat	5921	18.0 ± 10.9	0.151	0.210	0.616
Ex 16	0.01	Stirring under heat	359	32.6 ± 13.2	0.237	0.318	0.772
Ex 17	0.01	Ultrasonic dispersion	5300	16.8 ± 5.3	0.178	0.245	0.669
Ex 18	0.01	Ultrasonic dispersion	2211	18.1 ± 5.2	0.203	0.276	0.714
Ex 19	0.01	Ultrasonic dispersion	2229	16.1 ± 6.4	0.209	0.284	0.725
Ex 20	0.01	Ultrasonic dispersion	2327	17.0 ± 5.1	0.193	0.264	0.696
Ex 21	0.01	Ultrasonic dispersion	2386	34.6 ± 11.9	0.213	0.289	0.732
particle diameter: average \pm S.D.							

Example 22 to 26

In a glass beaker, 100 mg of the lipid A analog represented by the formula (III) was weighed. After addition of 50 mL of 0.003M NaOH, the resulting mixtures were stirred at $50 \pm 5^{\circ}\text{C}$ for 3, 8, 15, 30 and 90 minutes, respectively. To each of the reaction mixtures, 600 mL of a lactose phosphate buffer (an aqueous solution obtained by dissolving in 600 mL of distilled water for injection 100 g of lactose monohydrate, 0.45 g of $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$ and 0.35 g of $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$) was added. After stirring by a stirrer, a proper amount of distilled water for injection was added to adjust the total amount to 1 L. The medicament solution thus obtained was filtered through a $0.22 \mu\text{m}$ filter. Then, 5.3 ml portions of the filtrate were pipetted into vials, followed by freeze-drying. To each of the vials containing freeze-dried injection preparation, 5 ml of distilled water for injection was added to thaw the preparation. The evaluation results of the pharmacokinetics (AUC) of a rat, particle diameter, membrane fluidity (order parameter (S)) and surface charge are shown in Table 3. Since stirring was conducted in a sodium hydroxide solution at a temperature ($50 \pm 5^{\circ}\text{C}$) not less than the phase transfer temperature (about 30°C) of the lipid A analog represented by the formula (III), AUC depended on the stirring time and the longer the stirring time, the greater AUC. It has also been recognized that the greater AUC, the smaller order

parameter (S). Between AUC and the particle diameter or surface charge, no definite correlation was recognized.

Table 3

Influence of stirring time in sodium hydroxide on physicochemical parameters of an injection preparation containing a lipid A analog (500 μ g/vial) and on AUC (0-2hr) of a rat administered with the preparation

Example	Stirring time (min) in 0.003M NaOH	Dispersing method	AUC (0-2hr) (ng·hr/ml)	Particle diameter (nm)	Surface charge (zeta potential: mV)	Order parameter (S)
Ex 22	3	stirring under heat	1362	29.0 ± 14.0	0.215	0.759
Ex 23	8	stirring under heat	5668	18.4 ± 6.8	0.23	0.721
Ex 24	15	stirring under heat	7333	16.5 ± 6.0	0.242	0.695
Ex 25	30	stirring under heat	8051	15.3 ± 5.0	0.201	0.661
Ex 26	90	stirring under heat	9381	16.7 ± 5.3	0.151	0.636

particle diameter: Mean \pm S.D.

Example 27

In a glass beaker, 100 mg of the lipid A analog represented by the formula (III) was weighed, followed by the addition of 50mL of 0.003M NaOH. The resulting mixture was stirred by a stirrer at $50 \pm 5^{\circ}\text{C}$ for 30 minutes. To the reaction mixture was added 600 mL of a lactose phosphate (an aqueous solution obtained by dissolving in 600 mL of distilled water for injection 100 g of lactose monohydrate, 0.45 g of $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$ and 0.35 g of $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$). After stirring by a stirrer, a proper amount of distilled water for injection was added to adjust the total amount to 1 L. The resulting medicament solution was filtered through a $0.22\mu\text{m}$ filter. Each of 5.3 ml portions of the filtrate was pipetted into a vial and then freeze-dried. The initial stage product and products tested for stability under the conditions of 40°C and 75% RH (freeze-dried injection preparations stored for 1, 2 and 3 months) were thawed with 5 ml of distilled water for injection with the replicate of 3 and their membrane fluidity (order parameter (S)) was evaluated. The results are shown in Table 4. Variations in the order parameter (S) were small upon measurement, indicating that the reproducibility was sufficient. In addition, it was revealed that the injection preparations, whether they were initial stage products or stored for 3 months, were stable free from variations in order parameter (S) and pharmacokinetics.

Table 4

Change and reproducibility in order parameter (S) upon stability test of an injection preparation containing a lipid A analog (500 μ g/vial)

Measurement n	Initial	1M (40°C-75%R.H.)	2M (40°C-75%R.H.)	3M (40°C-75%R.H.)
1	0.560	0.589	0.572	0.597
2	0.586	0.565	0.589	0.576
3	0.599	0.599	0.591	0.591
Mean	0.584	0.584	0.588	0.588
CV(%)	3.40	2.99	1.78	1.84

Claims

1. A forecasting method of the pharmacokinetics of a lipid A analog in an injection preparation containing the lipid A analog or a pharmacologically acceptable salt thereof, which comprises measuring at least one of membrane fluidity and circular dichroism thereof.

2. The forecasting method according to claim 1, which is conducted for evaluating the injection preparation.

3. The forecasting method according to claim 1, which conducted for quality evaluation in order to obtain an injection preparation exhibiting constant pharmacokinetics.

4. The forecasting method according to claim 1, which is conducted in the preparation step of the injection preparation.

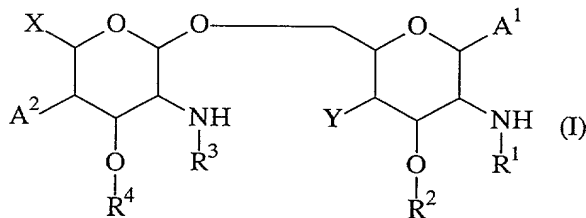
5. The forecasting method according to claim 1, wherein the membrane fluidity is measured by fluorescence probe method which uses, as parameters, at least one of order parameter (S), fluorescence polarity (P) and fluorescence anisotropy (r).

6. The forecasting method according to claim 1, wherein the injection, which contains aggregates having a diameter not greater than 30 nm, prepared by dissolving the lipid A analog or a pharmacologically acceptable salt

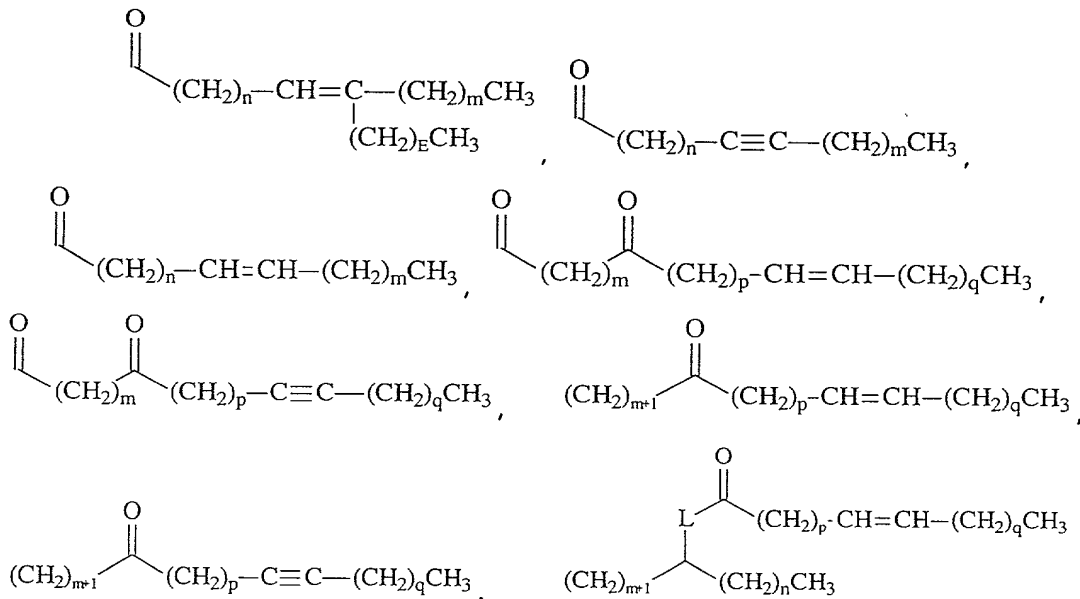
thereof in an alkaline aqueous solution and then adding a buffer thereto.

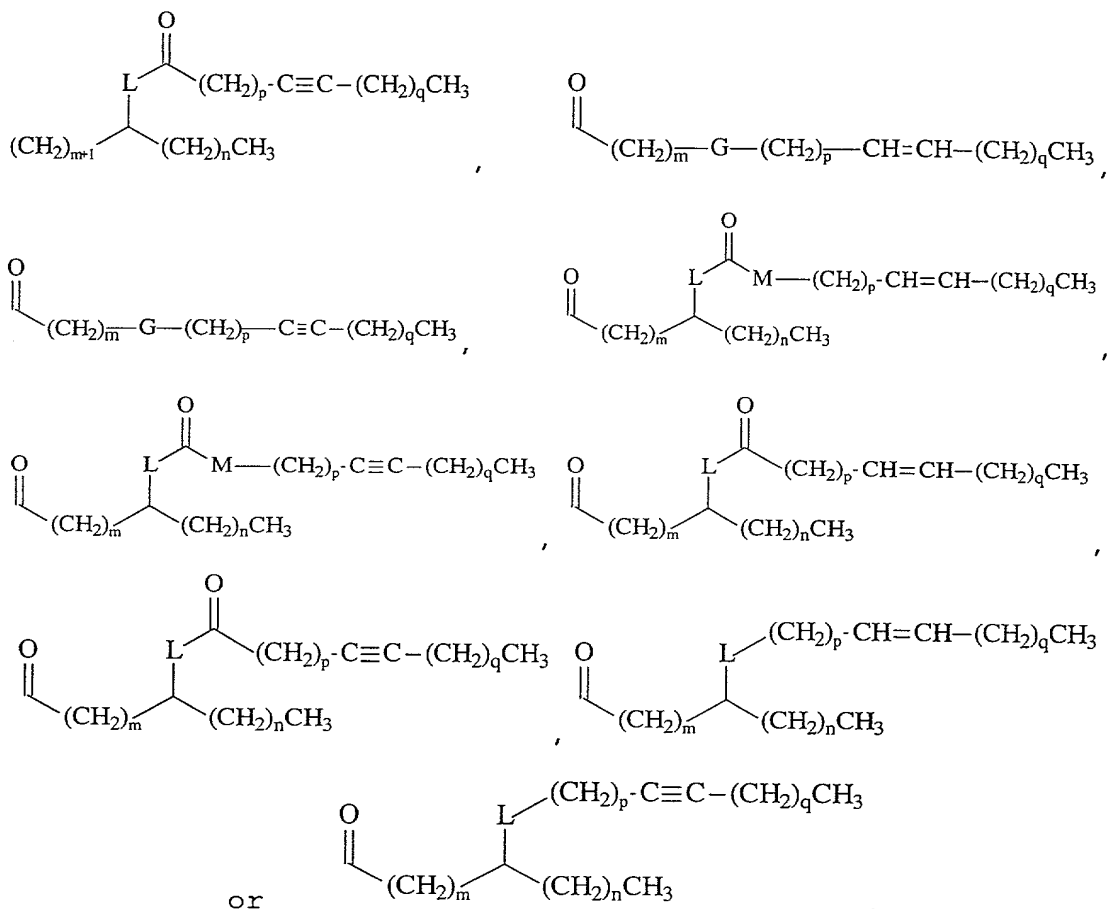
7. The forecasting method according to claim 1, wherein the injection preparation is an aqueous injection or freeze-dried preparation.

8. The forecasting method according to claim 1, wherein the lipid A analog or a pharmacologically acceptable salt thereof is a compound represented by the following formula (I):

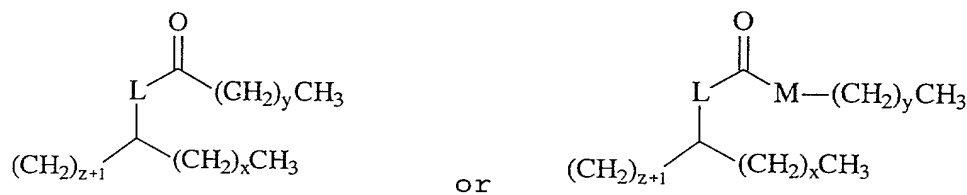
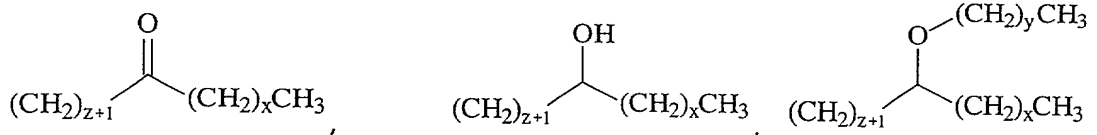
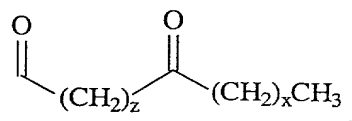
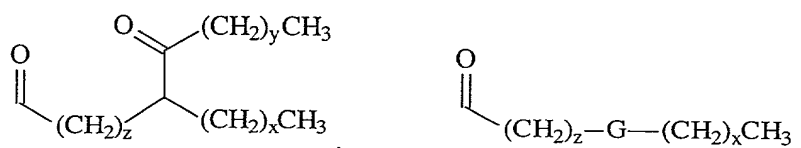
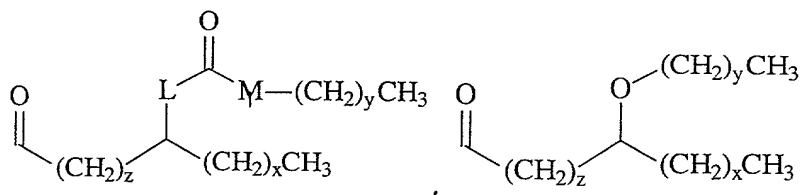
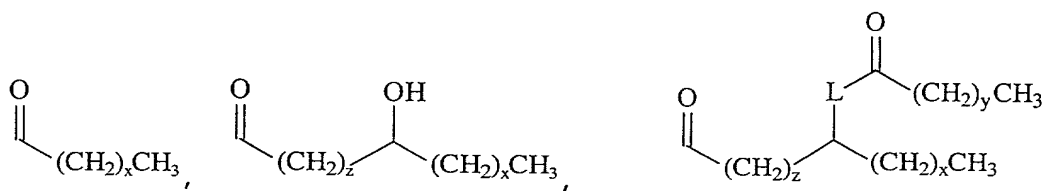


wherein at least one of R^1 , R^2 , R^3 and R^4 is



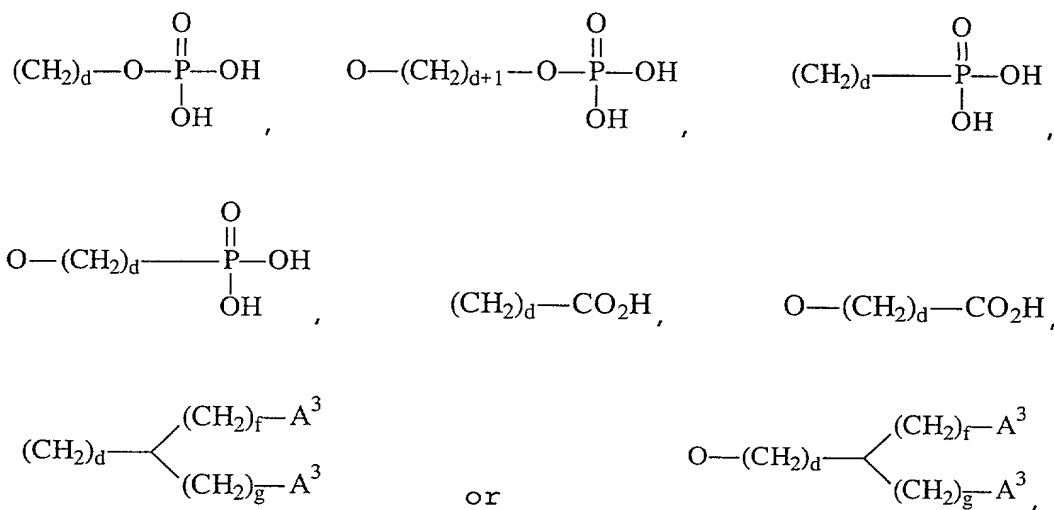


wherein each L is O, N or C; each M is O or N; each E independently is an integer of 0 to 14; each G independently is N, O, S, SO or SO₂; each m independently is an integer of 0 to 14; each n independently is an integer of 0 to 14; each p independently is an integer of 0 to 10; each q independently is an integer of 0 to 10, the rest of R¹, R², R³ and R⁴ are, independently of one another,

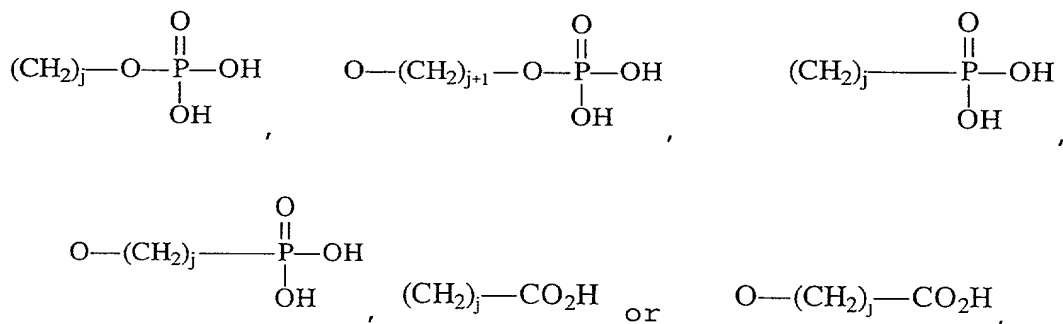


wherein each L is O, N or C; each M is O or N; each x independently is an integer of 0 to 14; each y independently is an integer of 0 to 14; each z independently is an integer of 0 to 10; each G independently is N, O, S, SO or SO₂.

A¹ and A² are, independently of one another, H, OH, OCH₃,



wherein each d independently is an integer of 0 to 5; each f independently is an integer of 0 to 5; each g independently is an integer of 0 to 5; each A³ independently is



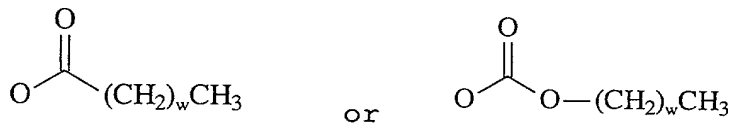
wherein each j independently is an integer of 0 to 14, X is H, (CH₂)_tCH₃, (CH₂)_tOH, (CH₂)_tO(CH₂)_vCH₃, (CH₂)_tOPO(OH)₂, (CH₂)_t-CH=CH-(CH₂)_vCH₃, (CH₂)_t-O-R⁵,



wherein t and v, are independently of one another, an

integer of 0 to 14; R^5 is any of the above definitions of R^1 to R^4 ,

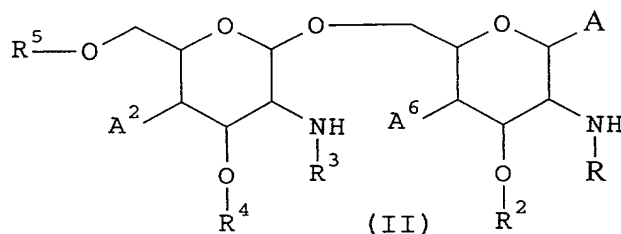
Y is H, OH, $O(CH_2)_wCH_3$, a halogen atom,



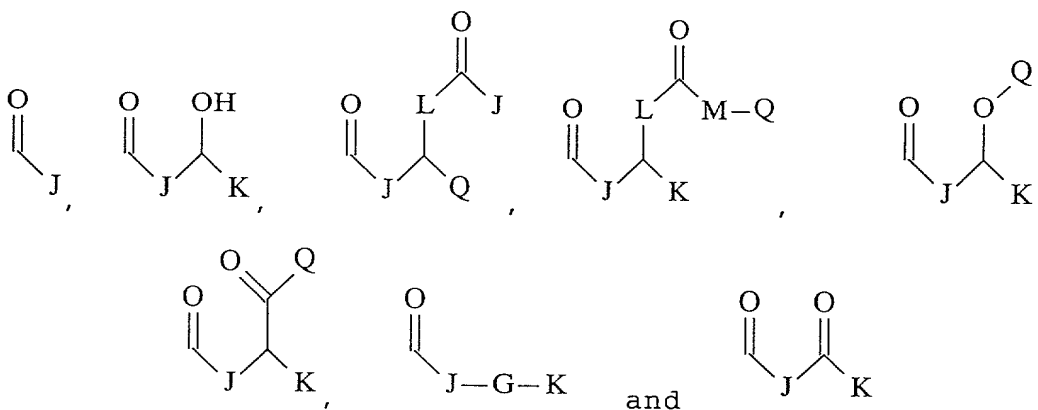
wherein w is an integer of 0 to 14,

or a pharmacologically acceptable salt thereof.

9. The forecasting method according to claim 1,
wherein the lipid A analog is a compound represented by the
following formula (II):



wherein R¹ is a group selected from the groups consisting of

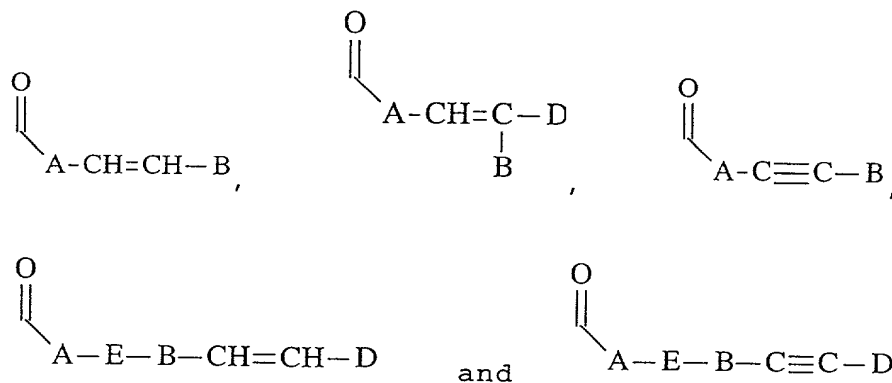


wherein J, K and Q are each a linear or branched alkyl group of 1 to 15 carbon atoms; L is O, NH₂ or CH₂; M is O or NH; G is NH, O, S, SO or SO₂,

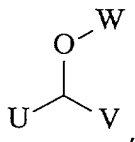
Figure 1 consists of 12 bar charts, each representing a different variable. The x-axis for all charts lists four groups: All respondents, Nonusers, Users, and Users with a computer at home. The y-axis represents the percentage of respondents, ranging from 0% to 100%.

- Age:** The distribution is relatively even across groups, with a slight peak for 'All respondents' in the 18-24 age range.
- Sex:** The distribution is relatively even across groups, with a slight peak for 'All respondents' in the 'Male' category.
- Education:** The 'Users' group shows a significantly higher percentage of respondents with a 'Bachelor's' degree compared to other groups.
- Income:** The 'Users' group shows a significantly higher percentage of respondents in the '\$30,000-\$49,999' income bracket compared to other groups.
- Computer use frequency:** The 'Users' group shows a significantly higher percentage of respondents who use the computer 'Daily' compared to other groups.
- Computer use location:** The 'Users' group shows a significantly higher percentage of respondents who use the computer 'At home' compared to other groups.
- Computer use purpose:** The 'Users' group shows a significantly higher percentage of respondents who use the computer for 'Work' compared to other groups.
- Computer use duration:** The 'Users' group shows a significantly higher percentage of respondents who use the computer for 'More than 1 year' compared to other groups.
- Computer use skill:** The 'Users' group shows a significantly higher percentage of respondents who are 'Very skilled' compared to other groups.
- Computer use confidence:** The 'Users' group shows a significantly higher percentage of respondents who are 'Very confident' compared to other groups.
- Computer use satisfaction:** The 'Users with a computer at home' group shows a significantly higher percentage of respondents who are 'Very satisfied' compared to other groups.
- Computer use anxiety:** The 'Users with a computer at home' group shows a significantly higher percentage of respondents who are 'Very anxious' compared to other groups.

R^2 is a linear or branched alkyl group of 5 to 15 carbon atoms, R^3 is a group selected from the groups consisting of



wherein E is N, O, S, SO or SO_2 ; A, B and D are each a linear or branched alkyl group of 1 to 15 carbon atoms, R^4 is a group selected from the groups consisting of a linear or branched alkyl group of 4 to 20 carbon atoms and

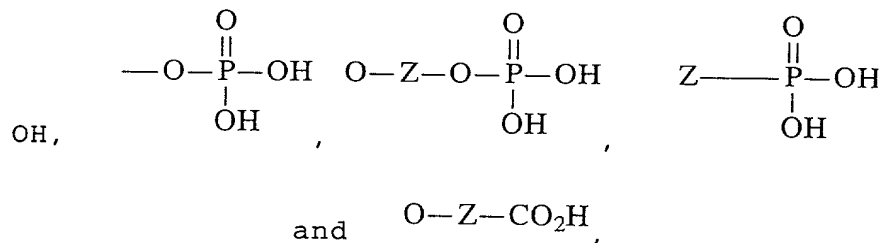


wherein U and V are each a linear or branched alkyl group of 2 to 15 carbon atoms; W is a hydrogen atom or a linear or branched alkyl group of 1 to 5 carbon atoms,

R^5 is a group selected from the groups consisting of a hydrogen atom, J' , $-J'-\text{OH}$, $-J'-\text{O}-K'$, $-J'-\text{O}-K'-\text{OH}$ and $-J'-\text{O}-\text{PO}(\text{OH})_2$, wherein J' and K' are each a linear or branched alkyl group of 1 to 5 carbon atoms,

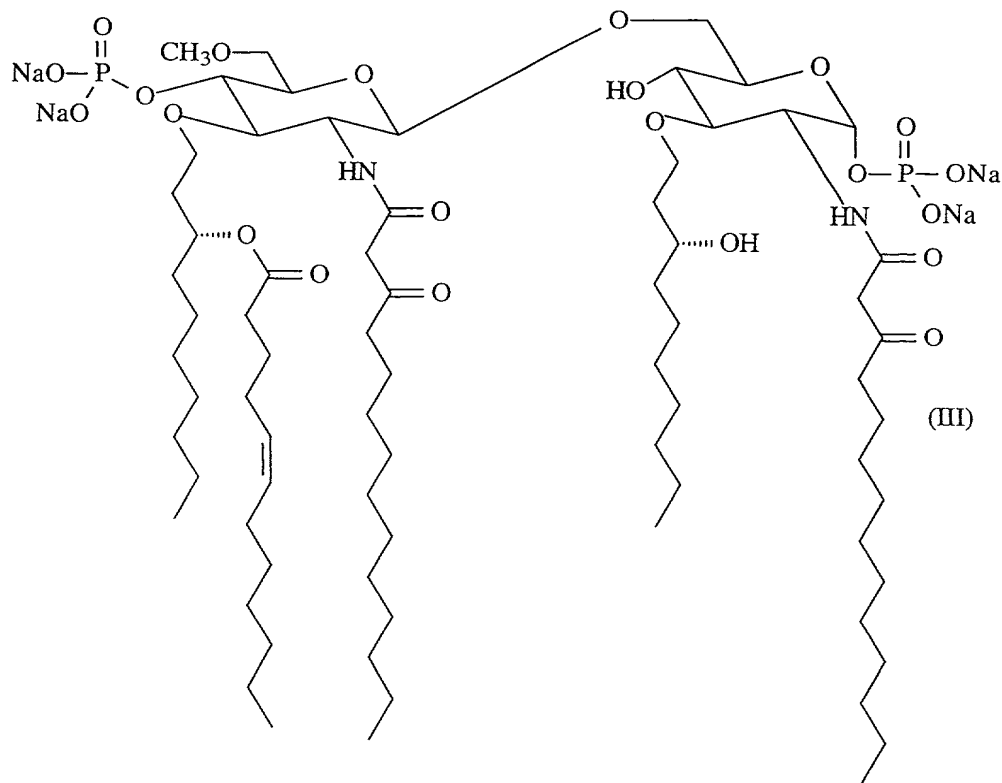
R^6 is a group selected from the groups consisting of a hydroxyl group, a halogen atom, an alkoxy group of 1 to 5 carbon atoms, and an acyloxy group of 1 to 5 carbon atoms,

A¹ and A² independently are each a group selected from the groups consisting of

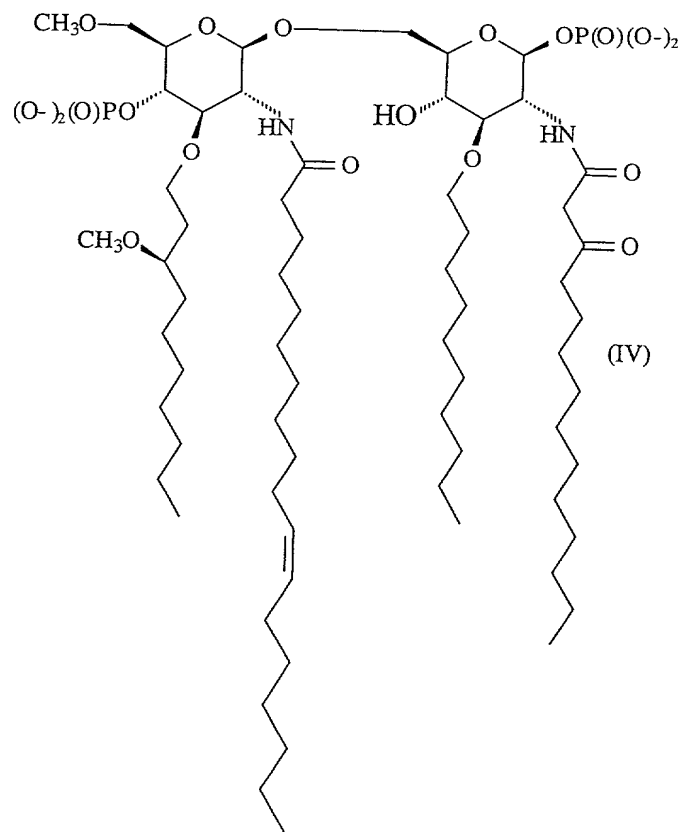


wherein Z is a linear or branched alkyl group of 1 to 10 carbon atoms,
or a pharmacologically acceptable salt thereof.

10. The forecasting method according to claim 1,
wherein the lipid A analog is a compound represented by the
following formula (III):



11. The forecasting method according to claim 1, wherein the lipid A analog is a compound represented by the following formula (IV):



12. The forecasting method according to claim 1, wherein the lipid A analog or a pharmacologically acceptable salt thereof has an aggregate structure in the form of endoplasmic reticulum with lipid biomolecular membrane or micelle.

Abstract

The present invention provides an evaluation method and preparation process of injection containing a lipid A analog. Specifically, it provides, in an injection preparation containing a lipid A analog or a pharmacologically acceptable salt thereof, forecasting and evaluating methods of the pharmacokinetics of the lipid A analog, and quality evaluating method and preparation process of the injection, which each comprises measuring membrane fluidity and/or circular dichroism.

Fig. 1

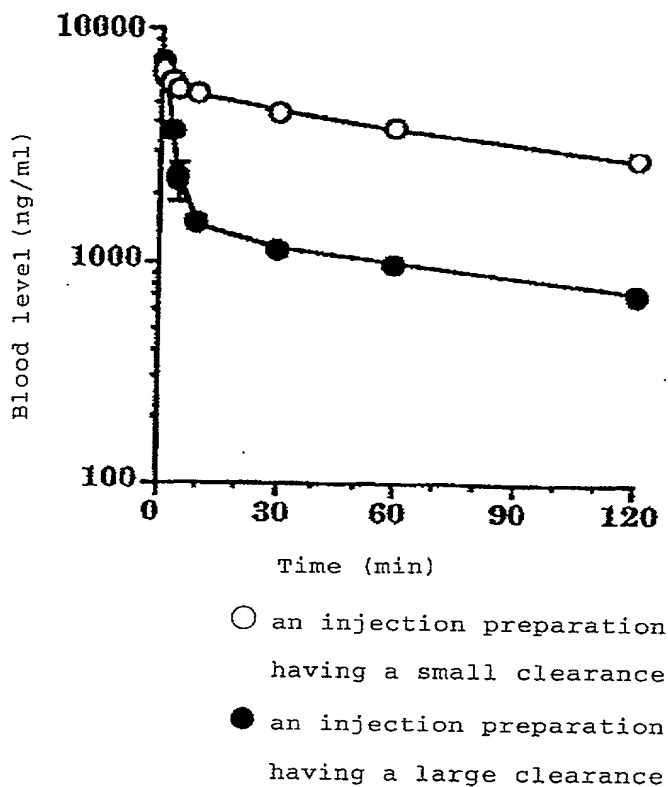


Fig. 2

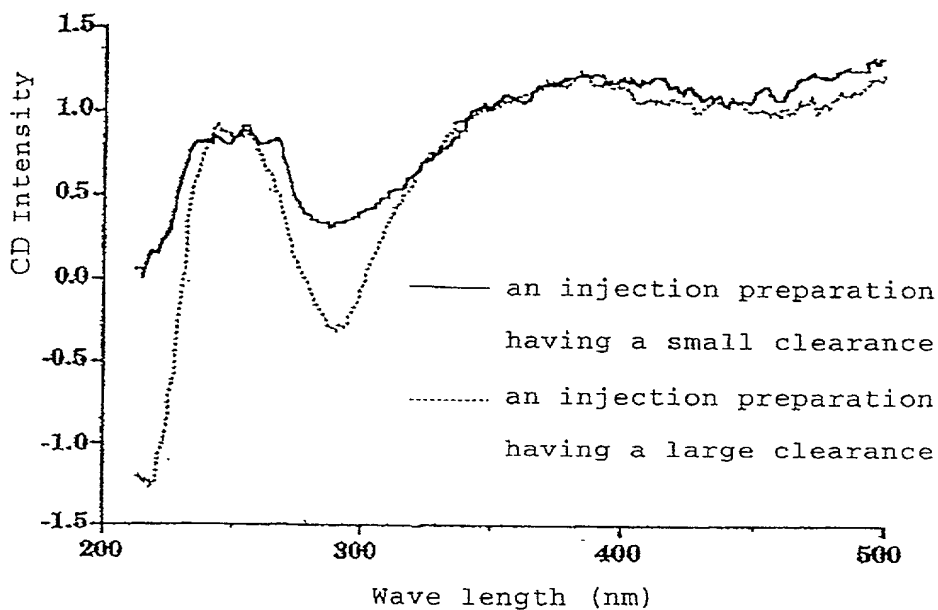


Fig. 3

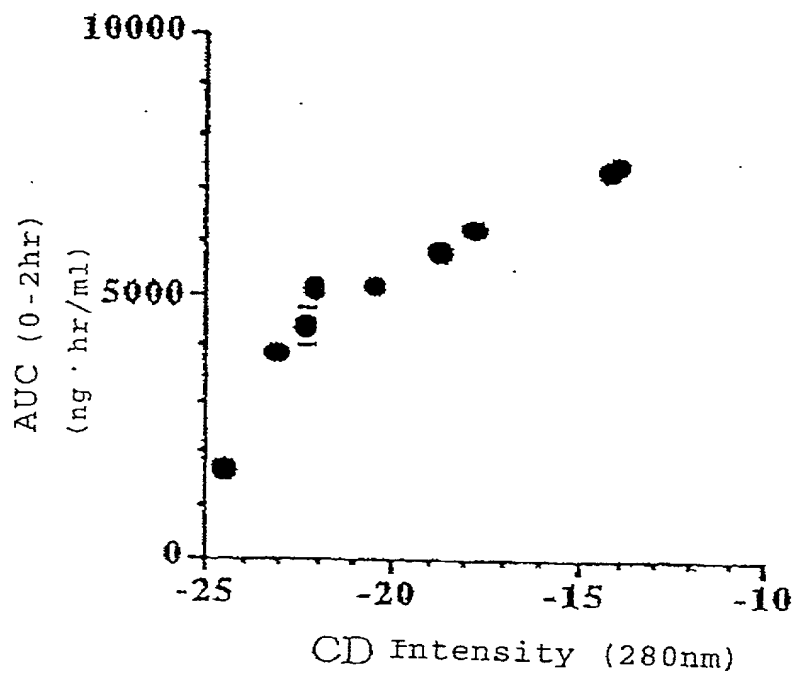


Fig. 4

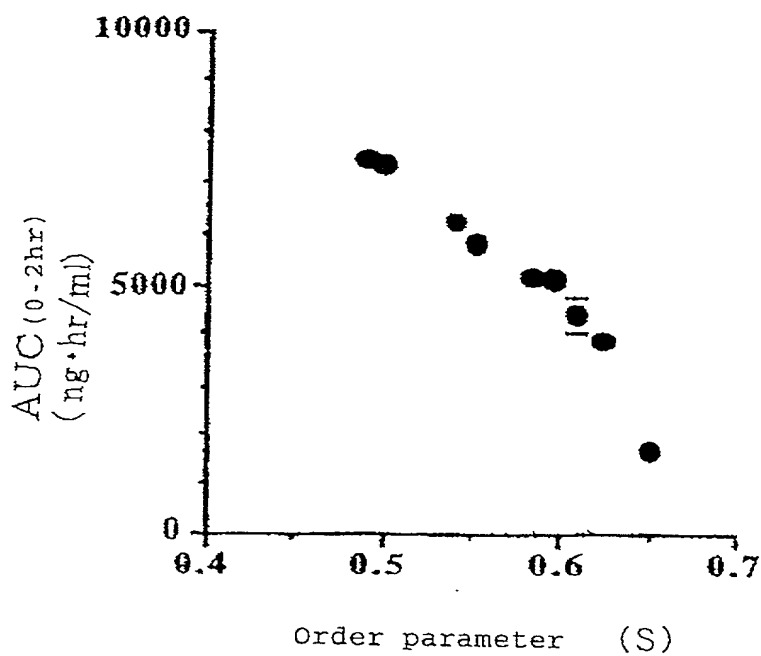


Fig. 5

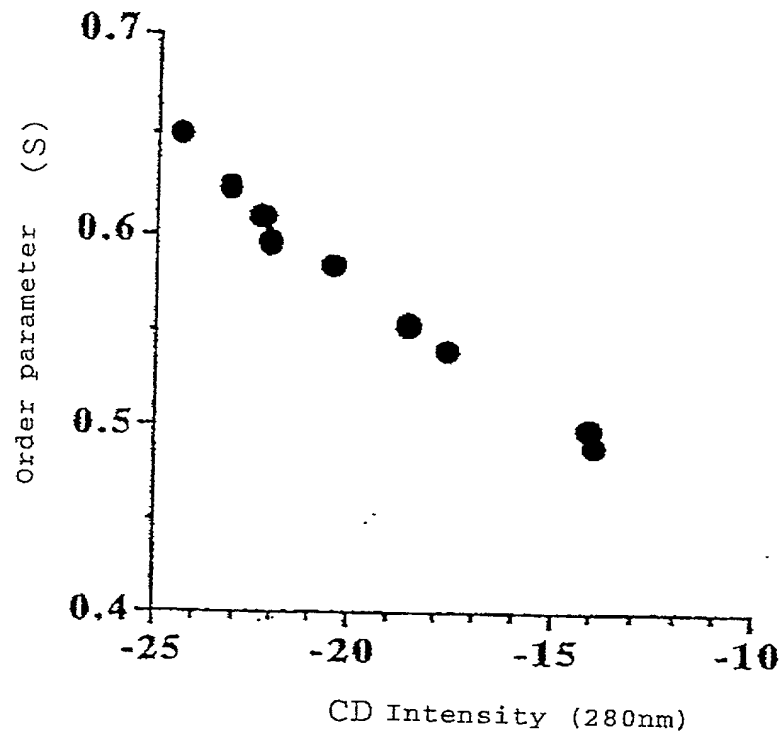


Fig. 6

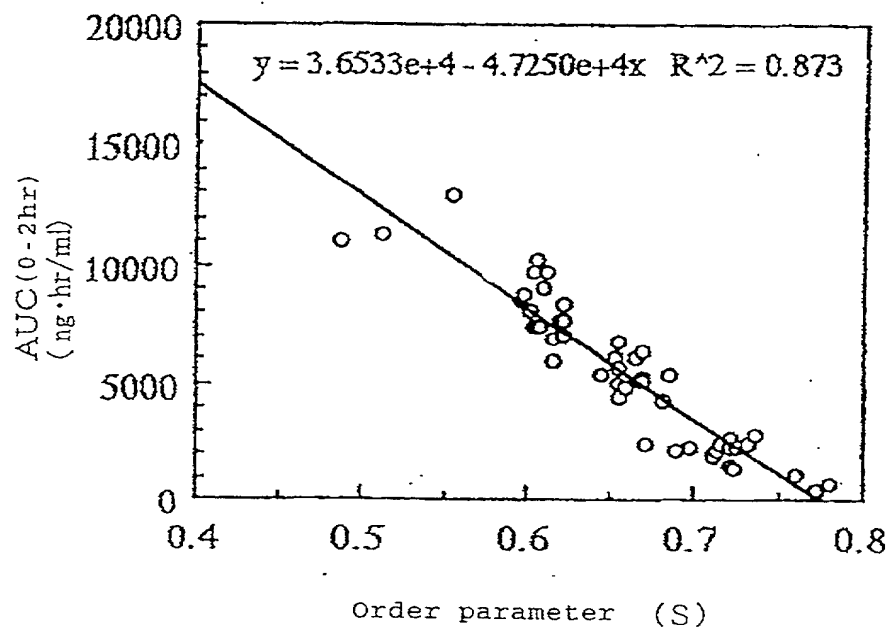


Fig. 7

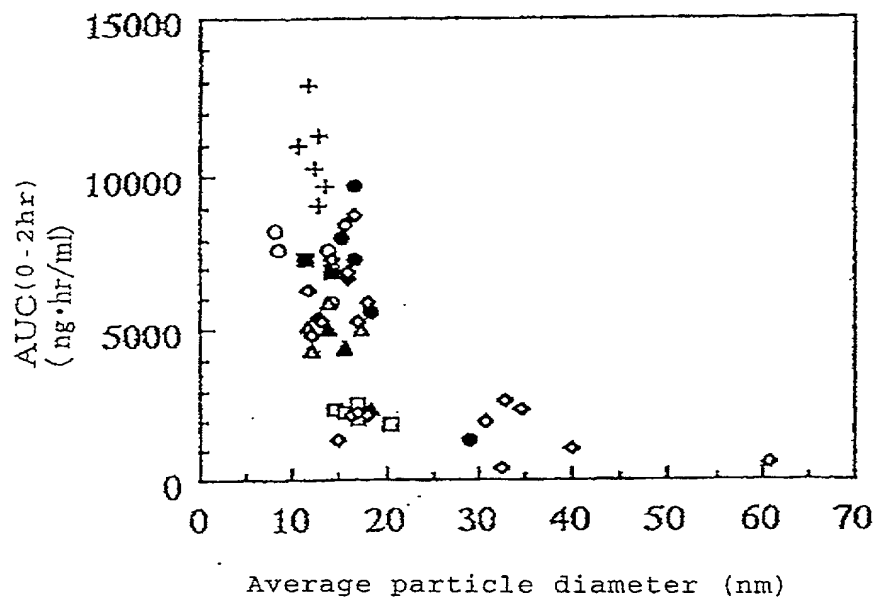
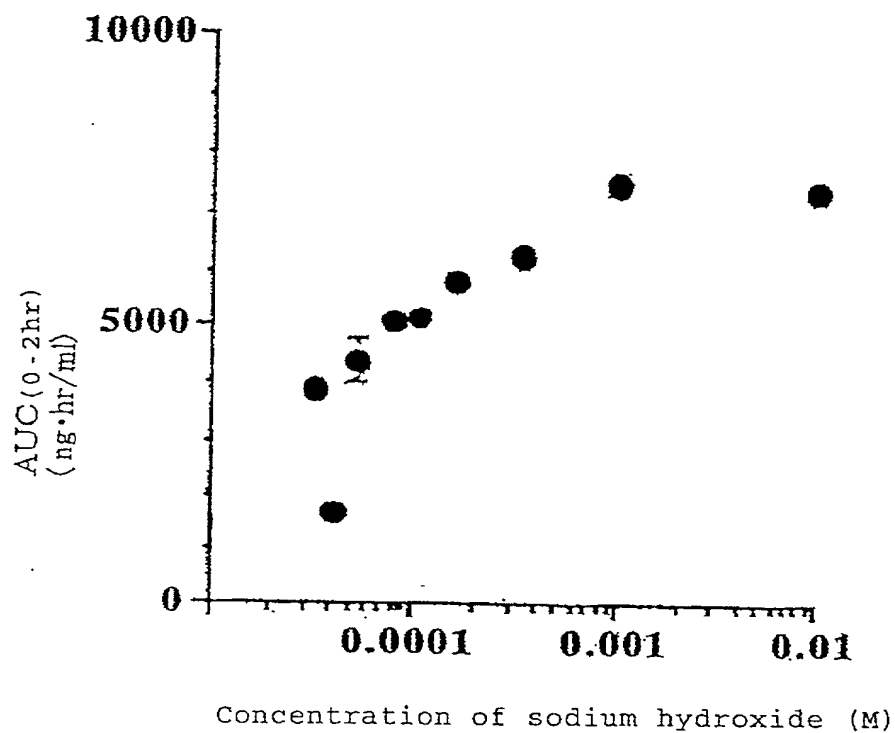


Fig. 8



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ATTORNEY DOCKET NO.

0425-0825P

PLEASE NOTE:
YOU MUST
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Insert Title:

As a below named inventor, I hereby declare that: my residence, post office address and citizenship are as stated next to my name; that I verily believe that I am the original, first and sole inventor (if only one inventor is named below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

Evaluation method and preparation process of injection
containing a lipid A analog

Fill in Appropriate
Information -
For Use Without
Specification
Attached:

the specification of which is attached hereto. If not attached hereto,

the specification was filed on _____ as
United States Application Number _____; and /or

the specification was filed on August 26, 1999 as PCT
International Application Number PCT/JP99/04615; and was
amended under PCT Article 19 on _____ (if applicable)

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.

I do not know and do not believe the same was ever known or used in the United States of America before my or our invention thereof, or patented or described in any printed publication in any country before my or our invention thereof or more than one year prior to this application, that the same was not in public use or on sale in the United States of America more than one year prior to this application, that the invention has not been patented or made the subject of an inventor's certificate issued before the date of this application in any country foreign to the United States of America on an application filed by me or my legal representatives or assigns more than twelve months (six months for designs) prior to this application, and that no application for patent or inventor's certificate on this invention has been filed in any country foreign to the United States of America prior to this application by me or my legal representatives or assigns, except as follows.

I hereby claim foreign priority benefits under Title 35, United States Code, §119 (a)-(d) of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

Insert Priority
Information:
(if appropriate)

Prior Foreign Application(s)

<u>10-246862</u> (Number)	<u>Japan</u> (Country)	<u>Sep. 1, 1998</u> (Month/Day/Year Filed)
_____ (Number)	_____ (Country)	_____ (Month/Day/Year Filed)
_____ (Number)	_____ (Country)	_____ (Month/Day/Year Filed)
_____ (Number)	_____ (Country)	_____ (Month/Day/Year Filed)
_____ (Number)	_____ (Country)	_____ (Month/Day/Year Filed)

Priority Claimed

<input checked="" type="checkbox"/> Yes	<input type="checkbox"/> No
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<input type="checkbox"/> Yes	<input type="checkbox"/> No
<input type="checkbox"/> Yes	<input type="checkbox"/> No
<input type="checkbox"/> Yes	<input type="checkbox"/> No

I hereby claim the benefit under Title 35, United States Code, §119(e) of any United States provisional application(s) listed below.

Insert Provisional
Application(s):
(if any)

_____ (Application Number)	_____ (Filing Date)
_____ (Application Number)	_____ (Filing Date)

All Foreign Applications, if any, for any Patent or Inventor's Certificate Filed More Than 12 Months (6 Months for Designs) Prior To The Filing Date of This Application:

Insert Requested
Information:
(if appropriate)

Country	Application No.	Date of Filing (Month/Day/Year)
_____	_____	_____
_____	_____	_____

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

Insert Prior U.S.
Application(s):
(if any)

_____ (Application Number)	_____ (Filing Date)	_____ (Status - patented, pending, abandoned)
_____ (Application Number)	_____ (Filing Date)	_____ (Status - patented, pending, abandoned)

I hereby appoint the following attorneys to prosecute this application and/or an international application based on this application and to transact all business in the Patent and Trademark Office connected therewith and in connection with the resulting patent based on instructions received from the entity who first sent the application papers to the attorneys identified below, unless the inventor(s) or assignee provides said attorneys with a written notice to the contrary:

Terrell C. Birch	(Reg. No. <u>19,382</u>)	Raymond C. Stewart	(Reg. No. <u>21,066</u>)
Joseph A. Kolasch	(Reg. No. <u>22,463</u>)	James M. Slattery	(Reg. No. <u>28,380</u>)
Bernard L. Sweeney	(Reg. No. <u>24,448</u>)	Michael K. Mutter	(Reg. No. <u>29,680</u>)
Charles Gorenstein	(Reg. No. <u>29,271</u>)	Gerald M. Murphy, Jr.	(Reg. No. <u>28,977</u>)
Leonard R. Svensson	(Reg. No. <u>30,330</u>)	Terry L. Clark	(Reg. No. <u>32,644</u>)
Andrew D. Meikle	(Reg. No. <u>32,868</u>)	Marc S. Weiner	(Reg. No. <u>32,181</u>)
Joe McKinney Muncy	(Reg. No. <u>32,334</u>)	Andrew F. Reish	(Reg. No. <u>33,443</u>)
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PLEASE NOTE:
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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full Name of First or Sole Inventor: Insert Name of Inventor Insert Date This Document is Signed Insert Residence Insert Citizenship Insert Post Office Address	GIVEN NAME	FAMILY NAME	INVENTOR'S SIGNATURE	DATE*
	<u>Kazuhiro KANEKO</u>		<u>Kazuhiro Kaneko</u>	Feb. 2, 2001
	Residence (City, State & Country)		CITIZENSHIP	
	<u>Ibaraki, Japan</u>		<u>Japanese</u>	
POST OFFICE ADDRESS (Complete Street Address including City, State & Country)				
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Full Name of Second Inventor, if any: see above	GIVEN NAME	FAMILY NAME	INVENTOR'S SIGNATURE	DATE*
	<u>Tomohiro WATANABE</u>		<u>Tomohiro Watanabe</u>	Feb. 2, 2001
	Residence (City, State & Country)		CITIZENSHIP	
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<u>Shatore KAWANAKA 403 3-7-9, Azuma, Tsukuba-shi, Ibaraki, Japan SPX</u>				
Full Name of Third Inventor, if any: see above	GIVEN NAME	FAMILY NAME	INVENTOR'S SIGNATURE	DATE*
	<u>Yasuyuki ASAI</u>		<u>Yasuyuki Asai</u>	Feb. 6, 2001
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	<u>Aichi, Japan</u>		<u>Japanese</u>	
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Full Name of Fourth Inventor, if any: see above	GIVEN NAME	FAMILY NAME	INVENTOR'S SIGNATURE	DATE*
	<u>Yoshihisa SANO</u>		<u>Yoshihisa Sano</u>	Feb. 2, 2001
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Full Name of Fifth Inventor, if any: see above	GIVEN NAME	FAMILY NAME	INVENTOR'S SIGNATURE	DATE*
	<u>Kiyomi KIKUCHI</u>		<u>Kiyomi Kikuchi</u>	Feb. 2, 2001
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<u>Green Corpo Fujishiro 2-512, 531, Miyawada, Fujishiro-machi, Kitasouma-gun, Ibaraki, Japan SPX</u>				

* DATE OF SIGNATURE

Full Name of Sixth
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Insert Name of Inventor
Insert Date This
Document Is Signed
Insert Residence
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GIVEN NAME FAMILY NAME INVENTOR'S SIGNATURE *DATE
Ikuo KUSHIDA Ikuo Kushida Feb.2, 2001

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Full Name of Seventh
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GIVEN NAME FAMILY NAME INVENTOR'S SIGNATURE *DATE
Kazuhide ASHIZAWA Kazuhide Ashizawa Feb.2, 2001

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Full Name of Eighth
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